# **Food Bioscience**

# The neglected nutrigenomics of milk: What is the role of inter-species transfer of small non-coding RNA? --Manuscript Draft--

Manuscript Number:	FBIO_2020_831R7				
Article Type:	Review Article				
Keywords:	Nutrigenomics; epigenetics; Milk; small non-coding RNA; nutrition; microbiome.				
Corresponding Author:	Laura Bordoni, Ph.D. Universita degli Studi di Camerino Camerino, ITALY				
First Author:	Laura Bordoni, Ph.D.				
Order of Authors:	Laura Bordoni, Ph.D.				
	Rosita Gabbianelli, Ph.D.				
Abstract:	The characterization of small non-coding RNA (sncRNA) in food has become part of the field as a promising field of nutrigenomics. Milk contains sncRNA that are protected by extracellular vesicles which makes them resistant to digestive processes and possibly absorbable by the human gut. Due to the high conservation of sncRNA, these molecules might mediate inter-species gene expression regulations, opening numerous applications in the field of human nutrition. These include the modulation of sncRNA milk profile through diet, both in humans and dairy animals, livestock rearing methods, food technology, but also the production of infant formulas or the usage of sncRNA as biomarkers. SncRNA contained in milk might contribute to the elucidation of the long-term effects of milk consumption in the human diet, confirming the application of nutrigenomics in both health promotion and food production areas. The main aim of this mini-review is to introduce this aspect of nutrigenomics illustrating both promising aspects and pitfalls.				
Suggested Reviewers:	Bodo Melnik melnik@t-online.de Expert in miRNA in milk				
	Patrick Provost patrick.provost@crchudequebec.ulaval.ca Espert in miRNA in milk				
	Gerd Schmitz ed.grubsneger-inu.kinilk@ztimhcs.dreg Espert in miRNA in milk				
Opposed Reviewers:					
Response to Reviewers:					



To Prof. Editors-in-Chief Jian Chen Jiangnan University, Wuxi, Jiangsu, China Joe Regenstein Cornell University, Ithaca, NY, United States Food Bioscience

Camerino, October 1st, 2020

Dear Editors,

we are pleased to resubmit the review article entitled *"The neglected nutrigenomics of milk: what is the role of inter-species transfer of small non-coding RNAs?"* by prof. Rosita Gabbianelli and me. We revised the manuscript according to the Editor's requests. Thank you for your efforts in improving the quality of this manuscript.

Yours Sincerely,

Laura Bordoni, PhD

Lauro Bordeni

The authors reviewed the manuscript according to the Editor's suggestions. All the issues have been accomplished.

1	HIGH	ILIGHTS
2	-	SncRNA are present in both plant- and animal-based food, especially in milk;
3	-	Exogenous sncRNA might regulate immune- and development-related processes;
4	-	Local effects and microbiome modulations are the major focus of sncRNA research;
5	-	Environment (i.e., diet) and health status modulate the milk's sncRNA profile;
6	-	SncRNA content might mediate nutrigenomic effects of milk in human nutrition.
7		
8		
9		
10		
11		
12		
13		
14		
15		
16		
17		
18		
19		
20		
21		
22		
23		
24		
25		

# 26 *Review*

# 27 The neglected nutrigenomics of milk: What is the role of inter-

# 28 species transfer of small non-coding RNA?

- 29 Laura Bordoni<sup>1\*</sup> and Rosita Gabbianelli<sup>1</sup>
- <sup>1</sup>School of Pharmacy, Unit of Molecular Biology, University of Camerino, Camerino, MC,
- 31 Italy
- 32
- 33 \*Corresponding author
- 34 Laura Bordoni
- 35 School of Pharmacy, Unit of Molecular Biology
- 36 University of Camerino
- 37 Via gentile III da Varano
- 38 62032, Camerino (MC), Italy
- 39 laura.bordoni@unicam.it
- 40 tel. 0737 403211
- 41
- 42 Running title: Nutrigenomics of dairy products: Focus on short non-coding RNA
- 43
- 44

#### 45 ABSTRACT

The characterization of small non-coding RNA (sncRNA) in food has become part of the 46 field as a promising field of nutrigenomics. Milk contains sncRNA that are protected by 47 extracellular vesicles which makes them resistant to digestive processes and possibly 48 absorbable by the human gut. Due to the high conservation of sncRNA, these molecules 49 50 might mediate inter-species gene expression regulations, opening numerous applications in the field of human nutrition. These include the modulation of sncRNA milk profile through 51 diet, both in humans and dairy animals, livestock rearing methods, food technology, but also 52 53 the production of infant formulas or the usage of sncRNA as biomarkers. SncRNA contained in milk might contribute to the elucidation of the long-term effects of milk consumption in 54 the human diet, confirming the application of nutrigenomics in both health promotion and 55 56 food production areas. The main aim of this mini-review is to introduce this aspect of nutrigenomics illustrating both promising aspects and pitfalls. 57

58 **Keywords**: nutrigenomics; epigenetics; milk; small non-coding RNA; nutrition; microbiome.

59

60 List of abbreviation	ıs
-------------------------	----

- 61 DNMT DNA methyl transferase
- 62 EV extracellular vesicles
- 63 miRNA micro RNA
- 64 mRNA messenger RNA
- 65 piRNA PIWI-interacting RNA
- 66 siRNA short interfering RNA

68	<b>F</b>	
69	Index	of contents
70	1.	Small non-coding RNA and gene regulation
71	2.	Inter-species gene expression regulation through small non-coding RNA: any
72		nutrigenomic effect?
73	3.	SncRNA in milk
74	4.	Concerns on the effects of exogenous sncRNA exposure
75	5.	Implications of milk sncRNA intake for human nutrition and future prospectives
76		5.1 Modulating the sncRNA profile for a healthy milk
77		5.2 SncRNA in infant formulas
78		5.3 Milk sncRNA as biomarkers
79		5.4 SncRNA profile in plant-based beverages used as milk substitutes
80	6.	Conclusions
81		

82 1. Small non-coding RNA and gene regulation

Small non-coding RNA (sncRNA) are untranslated transcripts (~21-34 nucleotides long) that 83 regulate 40 % to 60 % of gene expression in humans. Several types of sncRNA have been 84 described. MicroRNA (miRNA), endogenous short interfering RNA (endo-siRNA) and Piwi-85 interacting RNA (piRNA) are the most extensively studied. They differ in biogenesis, length, 86 and mechanisms through which they accomplish their biological functions (Carvalho 87 Barbosa, Calhoun & Wieden, 2020). The ability to downregulate gene expression in the 88 cytoplasm by pairing with target mRNA, mediated by the assembly of an effector complex, 89 i.e., RNA-induced silencing complex (RISC) (Bartel, 2004; Fabian & Sonenberg, 2012), is a 90

91 function of sncRNA (miRNA in particular), but it is not the only one. They can also act inside the nucleus (Sarshad et al., 2018), where they can post-transcriptionally regulate small 92 and long non-coding RNA or even promote gene expression at the transcriptional level 93 94 through mechanisms that have not been completely elucidated (Liu et al., 2018). Moreover, the specific subclass of piRNA can suppress the activities of transposable elements, regulate 95 chromatin architecture to control genomic stability and modulate stability and translation of 96 messenger RNA (Jodar & Anton, 2018). SncRNA have been found not only in the 97 intracellular environment (as initially hypothesized) but also in biological fluids. 98 99 Extracellular vesicles (EV), which are released as a means of intercellular communication, and fat globules have been shown to protect and carry sncRNA (van Herwijnen et al., 2018; 100 101 Wolf, Baier & Zempleni, 2015), thus facilitating the long-range intercellular effects of these 102 molecules (Ferrero et al., 2017; Turchinovich, Samatov, Tonevitsky & Burwinkel, 2013; Yeri et al., 2017). However, the presence of sncRNA in biological fluids is not exclusively 103 associated with EV (Turchinovich, Weiz, Langheinz & Burwinkel, 2011). Biological fluids 104 that contain sncRNA include blood, urine, saliva, cerebrospinal fluid, sperm, tears and milk 105 (Ferrero et al., 2017; Izumi et al., 2012; Park et al., 2009; Weber et al., 2010; Yeri et al., 106 107 2017). Their unexpected resistance to degradation in the extracellular environment led to sncRNA being considered strong biomarker of health and disease, as well as a promising 108 prognostic tools. Alterations in creations in creation of the second profile has been shown in numerous pathological 109 110 conditions, not only in the measurement of tissue-specific expression patterns (de Almeida, Fraczek, Parker, Delneri & O'Keefe, 2016; Lekka & Hall, 2018), but also in the detection of 111 peripheral sncRNA in body fluids (Gupta, Bang & Thum, 2010; Y. Jin et al., 2019; Mi, 112 113 Zhang, Zhang & Huang, 2013; Redell, Moore, Ward, Hergenroeder & Dash, 2010; Roth et al., 2010; Santamaria-Martos et al., 2019). 114

115

# 116 2. Inter-species gene expression regulation through small non-coding RNA: Any

117

### nutrigenomic effect?

Nutrigenomics studies how food can modulate gene expression (Bordoni & Gabbianelli, 118 2019). This discipline focuses on the role of macro- and micro-nutrients, bioactive 119 compounds and dietary regimens in regulating gene expression and consequentially affecting 120 121 the health status. In particular, nutri-epigenomics investigates the role of epigenetics in mediating the effects of food on gene expression. The term nutrimiromics has been coined to 122 define the study of how nutrients and bioactive molecules (e.g., selenium, zinc, resveratrol, 123 curcumin and quercetin) can modulate miRNA concentrations in the human body 124 (Quintanilha, Reis, Duarte, Cozzolino & Rogero, 2017). While the ability of food to modulate 125 endogenous sncRNA production has been extensively shown, the existence of food-derived 126 127 sncRNA, that remain stable and can potentially be absorbed, was shown by Yang, Hirschi and Farmer 2015 and it is still a discussed topic (Yang, Hirschi & Farmer, 2015). 128

129 Exogenous sncRNA have been found in both plant- and animal-derived foods. The mobility 130 of sncRNA from one species to another is considered one of the main mechanisms for crosstalk between different organisms, even between species from different kingdoms (Choi, Um, 131 Cho & Lee, 2017; Zeng et al., 2019). While the sequence of some miRNA is specific to a few 132 plants or animal lineages, others are conserved in animals and plants (Ha, Pang, Agarwal & 133 Chen, 2008; van Herwijnen et al., 2018). Since foods contain sncRNA that could potentially 134 target human genes, it has been speculated whether an inter-species genomic regulation by 135 sncRNA could exist (Li, Xu & Li, 2018; Liang et al., 2012; Zempleni, Baier, Howard & Cui, 136 2015; Zhang, Chen, Yin, Zhang & Zhang, 2019; Zhao, Cong & Lukiw, 2018) and have a 137 specific role in disease pathogenesis (Perge, Nagy, Decmann, Igaz & Igaz, 2017). However, 138 although sequence conservation of miRNA and target genes may suggest conservation of 139 expression patterns and functions, several questions remain to be addressed: the stability and 140

141 bioavailability of sncRNA as a function of the food matrix, the efficiency of their uptake in the gut system and the amount of xeno-microRNA needed for biological actions. Major 142 concerns about the possibility that sncRNA could have significant biological effects in 143 mammals have been raised (Dickinson et al., 2013; Snow, Hale, Isaacs, Baggish & Chan, 144 2013; Witwer, McAlexander, Queen & Adams, 2013). On the other hand, some authors have 145 described a significant bioavailability of both plants and animal-derived sncRNA (Baier, 146 147 Nguyen, Xie, Wood & Zempleni, 2014; Benmoussa et al., 2020; Manca et al., 2018; Wu et al., 2019), suggesting that they might be absorbed in the intestine and transferred into the 148 149 blood circulation (Liang et al., 2014; Liang et al., 2015; Yang, Farmer, Agyekum, Elbaz-Younes & Hirschi, 2015; Yang, Farmer, Agyekum & Hirschi, 2015; Zhao et al., 2018). Izumi 150 et al. (2015) showed that miRNA from milk might be taken up by human intestinal cells 151 152 (Liao, Du, Li & Lönnerdal, 2017) and macrophages (Izumi et al., 2015; Lässer et al., 2011). This suggested that certain types of food, beyond being a source of macro- or micro-nutrients, 153 bioactive molecules, and energy, might also provide biologically active sncRNA. Although 154 the possibility of systemic effects is still open (see paragraph 4 for more details), the exposure 155 to exogenous sncRNA coming from food has been reported (Ledda, Ottaggio, Izzotti, Sukkar 156 & Miele, 2020; Li et al., 2018; Sanchita, Trivedi, Asif & Trivedi, 2018; Vaucheret & 157 Chupeau, 2012), and the possibility that they might exert significant biological effects in 158 mammals needs further study (Asgari, 2017; Nguyen, 2020). 159

Bacteria could also produce miRNA-like molecules that could modulate the host's gene expression, as previously shown for sncRNA produced by viruses (Cardin & Borchert, 2017; Duval, Cossart & Lebreton, 2017; Kincaid & Sullivan, 2012; Shmaryahu, Carrasco & Valenzuela, 2014). However, only limited data is available on their ability to target human gene expression (Choi et al., 2017; Lee, 2019). On the other hand, bacteria manipulate the expression of various miRNA in the host to modulate cellular processes that favors their survival and proliferation (Ahmed, Zheng & Liu, 2016; Duval et al., 2017). Moreover, it has
been shown that fecal miRNA (including those deriving from food) can shape the gut
microbiota, thus representing a potential future strategy for manipulating the human
microbiome (Liu et al., 2016).

170

#### 171 3. sncRNA in milk

Milk is one of the most important biological fluids, rich in macro- and micro-nutrients but 172 173 also bioactive compounds, like antimicrobial molecules, growth factors, immune cells and antibodies. Moreover, milk is a rich source of all types of sncRNA (Martin, 2017; Testroet et 174 al., 2018; Weber et al., 2010), which are stable with degradative conditions (Izumi et al., 175 176 2012; Zempleni et al., 2016) and in vitro digestion (Benmoussa et al., 2016; Rani, Vashisht, et al., 2017). Their stability is favored by the presence in the emulsion of EV that protect 177 sncRNA from enzymatic degradation and facilitates their uptake by endocytosis (Baier et al., 178 2014; Pathan et al., 2019; Tomé-Carneiro et al., 2018; Zhou et al., 2012). Probably for this 179 reason, milk is the biological fluid that contains the highest level of sncRNA relative to its 180 181 volume, and these sncRNA are stable with acidic conditions, resistant to RNAse and to degradation with freeze-thaw cycles (Golan-Gerstl et al., 2017; Pieters et al., 2015; Rani, 182 Yenuganti, Shandilya, Onteru & Singh, 2017; Weber et al., 2010). 183

Despite numerous inter-species differences on sncRNA profile nave been measured, analysis of miRNA in milk from different species showed that some microRNA are persistently abundant and overlap between human and other mammal's milk. Benmoussa and Provost (2019) have provided a complete overview of the miRNA characterized in previous studies and identified the top 10 microRNA found in human, cow or goat milks. The existence of these recurrent "milk miRNA" suggests a conserved evolutionary process that leads to the 190 release of specific microRNA in milk, maybe because of potentially conserved functions in lactation and, possibly, for the newborn's development and health (Kosaka, Izumi, Sekine & 191 Ochiya, 2010; Stephen et al., 2020; van Herwijnen et al., 2018; Zempleni et al., 2016). It has 192 193 also been speculated that milk-derived sncRNA may be involved in the "epigenetic priming" of the newborn (Perge et al., 2017). Since the digestive tract of infants is far less developed 194 and has less harsh conditions (lower acidity and lower enzymatic activity), it allows immune 195 cells and, other cells in milk, to survive and settle within the infant's digestive tract wall (Le 196 Huërou-Luron, Blat & Boudry, 2010; Mirza et al., 2019). Thus, it has been hypothesized that 197 these specific conditions might led to the transfer of dietary microRNA through milk EV to 198 infants (Izumi et al., 2012; Kosaka et al., 2010; Zhou et al., 2012), having a role in regulating 199 200 their development. Carney et al. (2017) showed that the miRNA profile of breast milk from 201 mothers of premature infants differs from that of mothers of term infants, suggesting that 202 premature delivery might stimulate the secretion of a milk with a microRNA profile that may have adaptive functions for growth in premature infants (Carney et al., 2017). 203

The discovery of sncRNA in milk has raised the question about what is the function of these 204 regulatory elements in this biological fluid. Numerous miRNA released in milk originate 205 206 from epithelial cells (mammary gland cells), but the identification of abundant immunityrelated microRNA suggested that they can also be released in milk from immune cells. The 207 208 presence of miRNA in colostrum (Van Hese, Goossens, Vandaele & Opsomer, 2020) 209 suggested a potential role of miRNA as important regulators of both immune- and development-related processes (Alsaweed, Lai, Hartmann, Geddes & Kakulas, 2016a; Carney 210 et al., 2017; Kosaka et al., 2010; Q. Zhou et al., 2012). MiRNA can regulate B- and T-cell 211 212 differentiation and affect interleukin production of macrophages, and their role in modulating inflammation has been documented (Rebane & Akdis, 2013). Bovine milk EV and associated 213 miRNA have been shown to be bioavailable and to distribute among murine tissues, 214

accumulating in particular in the liver and, to a lesser extent, in the spleen of mice (Manca etal., 2018), supporting the possibility of systemic effects induced by exogenous sncRNA.

Among milk miRNA, miR-148a is one of the best studied. Its sequence is highly conserved 217 (it is identical between humans and cows), and it regulates DNMT1 and DNMT3 expression, 218 thus affecting epigenetic homeostasis of DNA methylation. MiR-148a has been shown to 219 220 have a role in the molecular mechanisms of oncogenesis (Li, Deng, Zeng & Peng, 2016). Both positive and negative associations with cancer have been proposed for miR-148a: some 221 authors addressed it as a risk factor (Melnik & Schmitz, 2019), while others suggested that it 222 223 might exert a protective effect against cancer in infants (Golan-Gerstl et al., 2017). MiR-148a may regulate food intake and adipogenesis (Melnik & Schmitz, 2017) and it could also affect 224 the development of the nervous system (Li et al., 2016). Another miRNA highly expressed 225 226 (particularly in cow's milk) and having biologically relevant functions is miR-21-5p. This miRNA regulates cell growth and proliferation (Kumarswamy, Volkmann & Thum, 2011), 227 and it has been defined as an oncomiR (Feng & Tsao, 2016). Another miRNA typical of 228 cow's milk and with high homology to the human sequence is miR-30d. This miRNA targets 229 the 3'-UTR of TP53 (an oncosuppressor gene) to down-regulate the tumor suppressor p53 230 231 protein levels, thus bringing into question the potential beneficial effects of milk in the long-232 term (Melnik, 2017; Melnik & Schmitz, 2019). These are just a few examples aimed to focus 233 on the heterogeneous sncRNA profile in milk, which is rich in miRNA that can have both 234 positive and negative effects on health (Svoronos, Engelman & Slack, 2016). MiRNA deregulation is typically found in cancer, with oncomiRs that are overexpressed, while tumor 235 suppressive miRNA are underexpressed in cancer cells. If it is confirmed that milk miRNA 236 237 can enter both normal and tumor cells and affect their biological functions (Golan-Gerstl et al., 2017), studies on sncRNA profiling might help to determine additional molecular 238 mechanisms through which potentially harmful effects of milk consumption might be 239

mediated. Considering that the long term safety of milk consumption is still debated due to its
potential implication on cancer promotion (Fraser et al., 2020; Jeyaraman et al., 2019; Lu et
al., 2016). Therefore, investigating the role of exogenous sncRNA might give further insights
(Pirim & Dogan, 2020). To better understand which pathways could be modulated by
miRNA contained in milk, several authors (Benmoussa & Provost, 2019; Golan-Gerstl et al.,
2017) extensively reviewed the implications for human health of the most conserved miRNA
in milk, considering also their bioavailability and bioaccessibility.

While an extensive characterization of the milk miRNA profile has been defined, little is 247 known about which endogenous siRNA and piRNA are present in milk. Considering that 248 endogenous siRNA and piRNA not only modulate gene expression but also affect genome 249 stability, more research is needed. This topic also warrants attention considering that EV have 250 251 been explored as nanodevices for the development of new therapeutic applications, and milk EV may be viable natural nano-carriers for the delivery of miRNA- and siRNA-based drugs 252 (Agil et al., 2019; Arntz et al., 2015). Since the interest towards these technologies is 253 increasing (Chakraborty, Sharma, Sharma, Doss & Lee, 2017; Galley & Besner, 2020; Gorji-254 Bahri, Hashemi & Moghimi, 2018), their application in different therapies might also benefit 255 256 from more research.

257

258

# 4. Concerns on the effects of exogenous sncRNA exposure

Important discrepancies on the biological effects of these exogenous sncRNA sources in humans have been identified in the scientific literature (Li et al., 2018; Zhang et al., 2019). A large part of the scientific community recommends caution in drawing definite conclusions because of potential fallacies. Issues include contamination, technical artifacts and confirmation bias (Fromm, Tosar, Lu, Halushka & Witwer, 2018; Heintz-Buschart et al.,

2018; Kang et al., 2017; Witwer, 2018; Yang, Hirschi, et al., 2015). Different methods and 264 their specific limitations have probably led to the discrepancies. The sensitivity of the 265 methods and the proper use of internal and external controls may also affect the reliability of 266 the results. The role of confounders in plasma miRNA analysis has also been raised by Wang 267 et al. (2018). The existence of significant similarities between bovine and human miRNA 268 sequences is believed to be responsible for false positive results in the detection of dietary 269 miRNA in human blood. Indeed, the existence of systemic effects of sncRNA is still argued. 270 Important issues in replicating the evidence to confirm the presence of high levels of 271 272 circulating miRNA in blood after milk consumption have been raised (Auerbach, Vyas, Li, Halushka & Witwer, 2016). While some studies reported that exogenous miRNA are active 273 274 in the recipient organisms (Zhang et al., 2012), and regulate gene expression at distant organ 275 sites, subsequent studies have been unable to confirm an active role of diet-derived miRNA in mammalian circulation or tissues (Auerbach et al., 2016; Kirchner, Buschmann, Paul & 276 Pfaffl, 2020; Title, Denzler & Stoffel, 2015; Title, Denzler & Stoffel, 2015; Witwer, 2014; 277 Witwer & Zhang, 2017; Zempleni, Baier & Hirschi, 2015). The hypothesis that a weaker or 278 absent gut barrier (i.e., at early stages of development or in gut disorders) could facilitate the 279 passage of RNA molecules has been recently investigated in animal models. Kirchner et al. 280 (2020) confirmed that a transfer of protein from maternal milk to the child's circulation exists 281 282 before gut closure, but they were not able to show an increased transfer of RNA molecules 283 with the same conditions in calves. An increased transfer of RNA was not measured in the presence of increased gut permeability in mice (Yang, Elbaz-Younes, Primo, Murungi & 284 Hirschi, 2018). 285

The hypothesis of a systemic role of milk sncRNA in systemic circulation remains to be studied (Fritz et al., 2016; Wang et al., 2018). However, their presence in food and consequentially in the gut, should be considered for the potential local effects. Indeed, food derived miRNA have been detected in feces and gastrointestinal mucosa (Link et al., 2019). Since miRNA play a major role in determining intestinal cell fate (Dalmasso et al., 2010), their uptake from colonocytes (Liao et al., 2017) and macrophages (Lässer et al., 2011) might exert significant effects on gut and intestinal immune systems that needs further investigations. For example, it has been shown that milk exosome and miRNA depletion exacerbates cecal inflammation in an animal model (Wu et al., 2019).

A substantial portion of EV in milk seems to escape absorption and enter the large intestine, 295 and given the previously described interplay between host and bacteria by sncRNA, it is 296 297 likely that sncRNA contained in milk may also modulate the human microbiome. Indeed, plant-derived exosomal microRNA have been shown to modulate the microbiome (Teng et 298 al., 2018), and alterations of the gut microbiota were measured after oral administration of 299 300 bovine milk-derived EV in mice, whose intestinal immunity was enhanced by the treatment (Tong et al., 2020). The ability of miRNA to modulate the microbiome has been shown for 301 miRNA contained in feces independently of their origin. Oral administration of synthetic 302 miRNA affects specific bacteria in the gut (Liu et al., 2016). Similarly, plant-derived 303 exosome-like nanoparticles that contain RNA were taken up by the gut microbiota, whose 304 305 composition was altered (Huang, Pham, Davis, Yu & Wang, 2020; Teng et al., 2018; Zhou, 306 Paz, Sadri, Fernando & Zempleni, 2019). It has also been shown that miRNA in human milk 307 differed between mothers supplemented with probiotics instead of placebo (Simpson et al., 308 2015). These results suggested that sncRNA contained in food may be used to manipulate the microbiome. 309

The possibility of modulating the sncRNA profile contained in food (including milk) opens the way to interesting future prospectives for the development of functional foods that might be active because of their optimized sncRNA content.

313

## 314 5. Implications of milk sncRNA intake for human nutrition and future prospectives

- 315
- 316

# 5.1 Modulating the sncRNA profile: From diet to food technology

SncRNA expression pattern in milk is influenced by several factors, such as maternal 317 nutrition and environmental exposures (Chen et al., 2017; Wang et al., 2016). The fat content 318 of the maternal diet appears to have a major effect on miRNA's expression in milk and in the 319 neonate (Van Hese et al., 2020). Since plant miRNA have been detected in human breast milk 320 (Lukasik, Brzozowska, Zielenkiewicz & Zielenkiewicz, 2017), maternal plant intake might 321 impact breast milk's sncRNA profile. Not only diet but also pathological conditions of the 322 323 mother might modulate the sncRNA profile of their milk. For example, breast milk-derived 324 EV from mothers with type 1 diabetes show aberrant levels of miRNA (Mirza et al., 2019). Since sncRNA of maternal milk might play a role in the development of the newborn's 325 immune system by shaping its microbiome (Le Doare, Holder, Bassett & Pannaraj, 2018), the 326 modulation of sncRNA induced by diseases or dietary regimens during lactation should be 327 taken into account. The immunological state of the mammary gland seems to affect miRNA 328 expression as well. This is true in humans, where alterations of sncRNA in milk have been 329 suggested as a biomarker of different pathologies (Ferrero et al., 2017; Kelleher et al., 2019; 330 Rebane & Akdis, 2013), but also in cows diagnosed with subclinical mastitis (Duval et al., 331 2017; Sun et al., 2015), that show alterations of their milk's miRNA profile. 332

333 Since the sncRNA profile in milk is complex, to achieve an optimized composition in terms 334 of these regulatory elements is an ambitious goal. Despite the presence of several negative 335 effects induced by some sncRNA, the complete removal of these molecules in milk might not 336 be the best solution (Golan-Gerstl et al., 2017; Wu et al., 2019). Ideally, the selective removal/enrichment of some sncRNA in milk might be the answer, but it still represents a
future prospective (Gessner et al., 2019; Nguyen, 2020). Here the authors review the
mechanisms that, currently or in the future, could be potentially applied to optimize sncRNA
profile in milk (Figure 1).

A strategy to modulate the sncRNA profile could be the application of some processes 341 regularly used in food technology to guarantee the microbiological safety of milk. Although 342 a limited loss of sncRNA during storage has been shown (Howard et al., 2015; Hozumi et al., 343 2012), raw milk and its fat derivatives are rich in miRNA (Melnik & Schmitz, 2017). This is 344 likely due to the high stability of EV at low pH, after boiling and after multiple freeze-345 thawing cycles (Pieters et al., 2015). Pasteurization (78°C) and homogenization of milk have 346 a minor effect on the miRNA profile (Golan-Gerstl et al., 2017). This treatment does not 347 affect the recovery of miR-148a, which has been detected at high level in pasteurized. 348 homogenized, and skim milk fractions. On the contrary, boiling (100°C) and ultra-heat 349 treatment (130°C) of milk significantly decrease the levels of milk miRNA. It has been 350 shown that EV are significantly reduced in fermented milk (Yu, Zhao, Sun & Li, 2017). A 351 reduction in miR-29b and miR-21 was measured after fermentation, suggesting that a general 352 353 loss of sncRNA is likely to occur after this process. This evidence suggests that fermented milk products, such as yoghurt, might exert different miRNA-dependent effects on human 354 355 health in comparison to pasteurized milk. Only one study investigated sncRNA in cheese (Oh 356 et al., 2017), concluding that it was not possible to correlate sncRNA profile with microbial communities present in the product. Replication studies in different kinds of samples are 357 warranted to clarify the role of sncRNA in cheese manufacturing. The majority of studies 358 359 analyzed a few candidate miRNA after different technological treatments, but did not provide a complete sncRNA profiling; thus, further studies on a complete sncRNA profiling in 360 different dairy products are warranted. 361

362 Different rearing conditions of lactating animals represent other potential factors that affect the milk's sncRNA profile. A different physiological status of the dairy cattle can modulate 363 the levels of miRNA secreted in milk. For example, the levels of lactogenic hormones (i.e., 364 365 prolactin) affect cellular and extracellular miR-148a expression in bovine epithelial breast cells (Muroya et al., 2016). Increased miR-148a expression (which is associated with a 366 decreased DNMT1's expression) is considered an important hallmark of high performance 367 368 dairy cows, that may secrete more miR-148a into their milk compared to regular cows. MiRNA profile is also altered by infections, such as mastitis induced by *Staphylococcus* 369 370 aureus or Escherichia coli pathogens (Cai et al., 2018; W. Jin et al., 2014; Sun et al., 2015). Thus, miR-142-5p and miR-223 have been suggested as biomarkers for the early detection of 371 372 bacterial infections in the mammary gland. Different miRNA profiles were detected in dairy cows fed with high- and low-quality forages (Wang et al., 2016). A modulation of miRNA 373 profile was detected in cows exposed to dietary supplementation with 5% linseed or 374 safflower oil, suggesting that miRNA implicated in lipid metabolism are differentially 375 376 regulated (Li et al., 2015). A high-fat diet during lactation was able to alter milk's miRNA profile in mice (Chen et al., 2017). The replacement of forage fiber with non-forage fiber 377 sources in dairy cow's diets changed the expression of milk's miRNA (Quan et al., 2020). 378 This evidence suggests that milk composition (including ncRNA profile) is responsive to 379 dietary manipulation and to animal rearing conditions, with direct implications for dairy 380 production. Moreover, sncRNA profiling might represent, in the future, a tool to monitor the 381 health and physiological status of dairy livestock. Finally, considering the effects of sncRNA 382 on bacteria strains, different sncRNA profile might impact also fermentation and dairy 383 production. 384

385

#### 386

5.2 SncRNA in children's formulas

387 Although breastfeeding is highly recommended, it may not always be possible. Thus, infant formulas represent an industrially produced substitute for infant consumption that attempts to 388 mimic the nutritional composition of breast milk as closely as possible (Martin, Ling & 389 390 Blackburn, 2016). Most formulas are based on cow's milk or soymilk and characterizing the levels of sncRNA expression in these products might help to improve the nutritional 391 adequacy of these foods (Stephen et al., 2020). The assessment of miR-148a-3p, one of the 392 most highly expressed miRNA in milk, showed significantly lower levels of this miRNA in 393 infant formula compared to human milk (Chen et al., 2010; Golan-Gerstl et al., 2017). This 394 395 preliminary data suggests that the total amount of sncRNA is depleted in infant formulas, that lack this archaic epigenetic regulatory signaling system. This might impact the early 396 397 metabolic programming and the immune system development in the newborns who cannot 398 benefit from the maternal lactation during early life. An extensive characterization of 399 maternal milk could help to identify the most abundant sncRNA and their functions, with the future aim to optimize the profile of these regulatory molecules. However, this is an 400 401 ambitious and complex challenge. SncRNA' concentration in milk might change during lactation, like other nutrients, according to the infant's needs (Carney et al., 2017; Lukasik et 402 al., 2017). This implies that the sncRNA profiling should be performed in different periods of 403 lactation and that supplementation should be consequentially time-dependent. Moreover, the 404 addition of synthetic miRNA in formulas might not have the same effect in newborns as 405 406 miRNA naturally present in milk. Indeed, transient transfection of chemically synthesized miRNA showed different behaviors than endogenous miRNA, suggesting that special caution 407 must be taken (Jin et al., 2015). As suggested by Golan-Gerstl et al. (2017), miRNA could be 408 isolated from animal sources, since about 90% of miRNA found in human milk are also 409 present in cow's and goat's milk; however, a wider characterization of the effects of sncRNA 410

411 in different periods of human life is required before that companies could considered the
412 possibility to use them as a supplement (Nguyen, 2020).

413

414 5.3 Milk sncRNA as biomarkers

Molecular targets that could be used to exactly measure the amount of consumed food are 415 wanted and metabolomics is a proliferative research field. The usage of sncRNA as 416 biomarkers of food intake has been suggested, but some concerns have been raised (Witwer 417 & Zhang, 2017). Firstry, a useful marker of intake reflects both the identity and the dose of 418 the source material; conversely, the sequence conservation of miRNA is incompatible with 419 420 discrimination of specific food sources. Moreover, there are still some uncertainties about the 421 linear correlation between miRNA abundance in the source material and their dietary absorption (Yang, Hirschi, et al., 2015). These concerns actually originated from studies 422 investigating plants' miRNA, but the same doubts can be extended to animal derived 423 sncRNA. Considering that low level of miRNA are present in body fluids, a thorough 424 sequencing, with consequential high costs, would be necessary to accomplish this goal with 425 426 confidence. For all these reasons, additional studies and technical implementations are needed to define a practical usage of sncRNA as biomarkers of food intake. Since it has been 427 shown that sncRNA profile varies depending on environmental stimuli and infections, these 428 429 sncRNA might help to identify unwanted environmental exposures, status of illness in cows (Ma, Tong, Ibeagha-Awemu & Zhao, 2019) or diseases in women (Kelleher et al., 2019). 430 This aspect represents a stimulating future prospective for this research field both in human 431 432 and veterinary medicine.

433

434

# 5.4 SncRNA profiles in plant-based beverages used as milk substitutes

435 A growing number of people are eliminating milk from their diet because of several side effects, first among others, lactose intolerance. The usage of "plant-based milks" such as 436 beverages based on soy, rice, oat and coconut is spreading. Since the presence of plant-437 438 derived miRNA in human breast milk has been shown, the different sncRNA profiles between animal-derived and plant-based milk might be a further aspect to be considered in 439 human nutrition. Treatment with plant's sncRNA has been demonstrated to systemically 440 reduce inflammation and prevent symptoms of multiple sclerosis in an experimental 441 autoimmune encephalomyelitis (EAE) mouse model (Cavalieri et al., 2016). This suggests 442 that exogenous sncRNA might significantly contribute to health promotion. Due to their 443 presence in breast milk, plant-derived molecules might have an impact not only in humans 444 445 that are directly fed with them but also in their progeny (Lukasik et al., 2017). Since there are no data on the stability of sncRNA in plant-based milks to date, further investigations are 446 necessary. Finally, since both bovine and plant infant formulae are produced (Tzifi, 447 Grammeniatis & Papadopoulos, 2014), research on milk's sncRNA might find further 448 449 applications in child nutrition, where sncRNA are likely to contribute to infant protection and development (Alsaweed, Lai, Hartmann, Geddes & Kakulas, 2016b). Measuring the effects 450 of milk and plant based formulas (also in relation to their sncRNA content) on the gut 451 microflora could be of particular interest. 452

453

#### 454 *6.* Conclusions

The characterization of sncRNA in food is an emerging research field of nutrigenomics. While a certain body of evidence is available for miRNA, few investigations have been done on siRNA or piRNA in food. Since not only gene expression regulation but also genomic stability is affected by sncRNA, further studies able to provide a complete profile of sncRNA in food are necessary. Since sncRNA are resistant to digestive processes, exogenous sncRNA

contained in food could be absorbed by the human gut. While systemic effects are still 460 debated, it appears likely that they can affect the gut and the resident's microbiome. These 461 bioactive molecules could contribute to the impact of food on gene expression regulation and 462 their impact on human health. These gene regulation pathways represent a bridge between 463 different animal species, and between the animal and the plant kingdoms. However, a 464 scientific consensus on this topic is still missing. Publication bias (e.g., avoiding publishing 465 466 negative results), might contribute to these uncertainties. Clarifying the biological effects of sncRNA contained in milk could provide a complete overview on the effects of milk 467 468 consumption in human diet, since milk is a good source of nutrients but the full safety (concerning complex environmentally-based diseases) of its intake in the long term is still 469 470 discussed (Fraser et al., 2020; Jeyaraman et al., 2019; Lu et al., 2016). For this reason, 471 additional studies on this topic are warranted. These would help to clarify the whole picture 472 and to identify practical applications of this research field, that range from food technology, to animal rearing or infant formulas production. These applications are directed towards an 473 474 optimized molecular nutrition, promoting the role of molecular biology (and nutrigenomics in particular) beyond basic research. 475

476

- 477 **Conflicts of interest**
- 478 The authors declare no conflicts of interests.

479

- 480 Acknowledgements
- 481 The authors thank Francesca Buonanno for helping in the production of Figure 1.

482

483 (	Contri	butions

LB wrote the manuscript, RG revised and supervised the work. All authors have approved thefinal article.

486

## 487 Fundings

488 This research did not receive any specific grant from funding agencies in the public,489 commercial, or not-for-profit sectors.

490

### 491 **REFERENCES**

492 Ahmed, W., Zheng, K. & Liu, Z.F. (2016). Small non-coding RNA: New insights in

493 modulation of host immune response by intracellular bacterial pathogens. *Frontiers in* 

494 *Immunology; 7, 431*. doi:10.3389/fimmu.2016.00431

- 495 Alsaweed, M., Lai, C. T., Hartmann, P. E., Geddes, D. T. & Kakulas, F. (2016a). Human
- 496 milk cells and lipids conserve numerous known and novel miRNA, some of which are
- differentially expressed during lactation. *PloS One*, *11*(4), e0152610.
- 498 https://doi.org/10.1371/journal.pone.0152610
- 499 Alsaweed, M., Lai, C. T., Hartmann, P. E., Geddes, D. T. & Kakulas, F. (2016b). Human
- 500 milk miRNA primarily originate from the mammary gland resulting in unique miRNA
- 501 profiles of fractionated milk. *Scientific Reports*, *6*, 20680.
- 502 https://doi.org/10.1038/srep20680
- 503 Aqil, F., Munagala, R., Jeyabalan, J., Agrawal, A. K., Kyakulaga, A.-H., Wilcher, S. A. &
- 504 Gupta, R. C. (2019). Milk exosomes Natural nanoparticles for siRNA delivery. *Cancer*

- 505 *Letters*, 449, 186–195. https://doi.org/https://doi.org/10.1016/j.canlet.2019.02.011
- 506 Arntz, O. J., Pieters, B. C. H., Oliveira, M. C., Broeren, M. G. A., Bennink, M. B., de Vries,
- 507 M., van Lent, P., Koenders. M. K., van den Berg. W. B., van der Kraan, P. M. & van de
- 508 Loo, F. A. J. (2015). Oral administration of bovine milk derived extracellular vesicles
- attenuates arthritis in two mouse models. *Molecular Nutrition & Food Research*, 59(9),
- 510 1701–1712. https://doi.org/10.1002/mnfr.201500222
- 511 Asgari, S. (2017). RNA as a means of inter-species communication and manipulation:
- 512 Progresses and shortfalls. *RNA Biology*, 14, 389-390.
- 513 https://doi.org/10.1080/15476286.2017.1306172
- 514 Auerbach, A., Vyas, G., Li, A., Halushka, M. & Witwer, K. (2016). Uptake of dietary milk
- 515 miRNA by adult humans: A validation study. *F1000Research*, *5*, 721.
- 516 https://doi.org/10.12688/f1000research.8548.1
- 517 Baier, S. R., Nguyen, C., Xie, F., Wood, J. R. & Zempleni, J. (2014). MicroRNA are
- absorbed in biologically meaningful amounts from nutritionally relevant doses of cow
- 519 milk and affect gene expression in peripheral blood mononuclear cells, HEK-293 kidney
- 520 cell cultures, and mouse livers. *Journal of Nutrition*, *144*(10), 1495–1500.
- 521 https://doi.org/10.3945/jn.114.196436
- 522 Bartel, D. P. (2004). MicroRNA: Genomics, biogenesis, mechanism, and function. Cell,
- 523 *116*(2), 281–297. https://doi.org/10.1016/s0092-8674(04)00045-5
- 524 Benmoussa, A., Laugier, J., Beauparlant, C. J., Lambert, M., Droit, A. & Provost, P. (2020).
- 525 Complexity of the microRNA transcriptome of cow milk and milk-derived extracellular
- vesicles isolated via differential ultracentrifugation. *Journal of Dairy Science*, 103(1),
- 527 16–29. https://doi.org/10.3168/jds.2019-16880

22

528	Benmoussa, A., Lee, C. H. C., Laffont, B., Savard, P., Laugier, J., Boilard, E. & Provost, P.
529	(2016). Commercial dairy cow milk microRNA resist digestion under simulated
530	gastrointestinal tract conditions. <i>Journal of Nutrition</i> , 146(11), 2206–2215.
531	https://doi.org/10.3945/jn.116.237651
532	Benmoussa, A. & Provost, P. (2019). Milk microRNA in health and disease. Comprehensive
533	Reviews in Food Science and Food Safety, 18(3), 703–722.

https://doi.org/10.1111/1541-4337.12424 534

- 535 Bordoni, L. & Gabbianelli, R. (2019). Primers on nutrigenetics and nutri(epi)genomics:
- Origins and development of precision nutrition. *Biochimie*, 160, 156–171. 536
- https://doi.org/10.1016/j.biochi.2019.03.006 537
- 538 Cai, M., He, H., Jia, X., Chen, S., Wang, J., Shi, Y. & Lai, S. (2018). Genome-wide
- microRNA profiling of bovine milk-derived exosomes infected with Staphylococcus 539

aureus. Cell Stress & Chaperones, 23(4), 663-672. https://doi.org/10.1007/s12192-018-540

0876-3 541

- Cardin, S.-E. & Borchert, G. M. (2017). Viral microRNA, host microRNA regulating viruses, 542
- 543 and bacterial microRNA-like RNA. Methods in Molecular Biology (Clifton, N.J.), 1617,

39-56. https://doi.org/10.1007/978-1-4939-7046-9 3 544

Carney, M. C., Tarasiuk, A., DiAngelo, S. L., Silveyra, P., Podany, A., Birch, L. L., Paul, I. 545

M., Kelleher, S. & Hicks, S. D. (2017). Metabolism-related microRNA in maternal 546

- breast milk are influenced by premature delivery. Pediatric Research, 82(2), 226-236. 547
- 548 https://doi.org/10.1038/pr.2017.54
- Carvalho Barbosa, C., Calhoun, S. H. & Wieden, H.-J. (2020). Non-coding RNA: What are 549
- we missing? Biochemistry and Cell Biology, 98(1), 23-30. https://doi.org/10.1139/bcb-550

2019-0037 551

552	Cavalieri, D.	, Rizzetto, L.,	Tocci, N.,	Rivero,	D., Asquini	, E.,	, Si-Ammour,	А.,	Bonechi,	Е.,
-----	---------------	-----------------	------------	---------	-------------	-------	--------------	-----	----------	-----

553 Ballerini, C. & Viola, R. (2016). Plant microRNA as novel immunomodulatory agents.

554 Scientific Reports, 6, 25761. https://doi.org/10.1038/srep25761

- 555 Chakraborty, C., Sharma, A. R., Sharma, G., Doss, C. G. P. & Lee, S.S. (2017). Therapeutic
- 556 miRNA and siRNA: Moving from bench to clinic as next generation medicine.
- 557 *Molecular Therapy Nucleic Acids*, 8, 132–143.
- 558 https://doi.org/10.1016/j.omtn.2017.06.005
- 559 Chen, X., Gao, C., Li, H., Huang, L., Sun, Q., Dong, Y., Tian, C., Gao, S., Dong, H., Guan,
- 560 D., Hu, X., Zhao, S., Li, L., Zhu, L., Yan, Q., Zhang, J., Zen, K., Zhang, C.Y. & Zhang,
- 561 C.Y. (2010). Identification and characterization of microRNA in raw milk during
- 562 different periods of lactation, commercial fluid, and powdered milk products. *Cell*

563 *Research*, 20(10), 1128–1137. https://doi.org/10.1038/cr.2010.80

- Chen, Y., Wang, J., Yang, S., Utturkar, S., Crodian, J., Cummings, S., Thimmapuram, J., San
- 565 Miguel, P., Kuang, S., Gribskov, M., Plaut, K. & Casey, T. (2017). Effect of high-fat
- diet on secreted milk transcriptome in midlactation mice. *Physiological Genomics*,

567 *49*(12), 747–762. https://doi.org/10.1152/physiolgenomics.00080.2017

- 568 Choi, J. W., Um, J. H., Cho, J. H. & Lee, H. J. (2017). Tiny RNA and their voyage via
- 569 extracellular vesicles: Secretion of bacterial small RNA and eukaryotic microRNA.
- 570 *Experimental Biology and Medicine*, 242(15), 1475–1481.
- 571 https://doi.org/10.1177/1535370217723166
- 572 Dalmasso, G., Nguyen, H. T. T., Yan, Y., Laroui, H., Srinivasan, S., Sitaraman, S. V &
- 573 Merlin, D. (2010). MicroRNA determine human intestinal epithelial cell fate.
- 574 *Differentiation; Research in Biological Diversity*, 80(2–3), 147–154.
- 575 https://doi.org/10.1016/j.diff.2010.06.005

576	de Almeida, R. A., Fraczek, M. G., Parker, S., Delneri, D. & O'Keefe, R. T. (2016). Non-
577	coding RNA and disease: The classical ncRNA make a comeback. Biochemical Society
578	Transactions, 44(4), 1073-1078. https://doi.org/10.1042/BST20160089
579	Dickinson, B., Zhang, Y., Petrick, J. S., Heck, G., Ivashuta, S. & Marshall, W. S. (2013).
580	Lack of detectable oral bioavailability of plant microRNA after feeding in mice. Nature
581	Biotechnology, 31(11), 965–967. https://doi.org/10.1038/nbt.2737
582	Duval, M., Cossart, P. & Lebreton, A. (2017). Mammalian microRNA and long noncoding
583	RNA in the host-bacterial pathogen crosstalk. Seminars in Cell & Developmental
584	Biology, 65, 11-19. https://doi.org/10.1016/J.SEMCDB.2016.06.016
585	Fabian, M. R. & Sonenberg, N. (2012). The mechanics of miRNA-mediated gene silencing:
586	A look under the hood of miRISC. Nature Structural & Molecular Biology, 19(6), 586-
587	593. https://doi.org/10.1038/nsmb.2296
588	Feng, Y.H. & Tsao, C.J. (2016). Emerging role of microRNA-21 in cancer. Biomedical
588 589	Feng, Y.H. & Tsao, C.J. (2016). Emerging role of microRNA-21 in cancer. <i>Biomedical Reports</i> , 5(4), 395–402. https://doi.org/10.3892/br.2016.747
589	Reports, 5(4), 395-402. https://doi.org/10.3892/br.2016.747
589 590	Reports, 5(4), 395–402. https://doi.org/10.3892/br.2016.747 Ferrero, G., Cordero, F., Tarallo, S., Arigoni, M., Riccardo, F., Gallo, G. & Naccarati, A.
589 590 591	<ul> <li><i>Reports</i>, 5(4), 395–402. https://doi.org/10.3892/br.2016.747</li> <li>Ferrero, G., Cordero, F., Tarallo, S., Arigoni, M., Riccardo, F., Gallo, G. &amp; Naccarati, A. (2017). Small non-coding RNA profiling in human biofluids and surrogate tissues from</li> </ul>
589 590 591 592	<ul> <li><i>Reports</i>, 5(4), 395–402. https://doi.org/10.3892/br.2016.747</li> <li>Ferrero, G., Cordero, F., Tarallo, S., Arigoni, M., Riccardo, F., Gallo, G. &amp; Naccarati, A. (2017). Small non-coding RNA profiling in human biofluids and surrogate tissues from healthy individuals: Description of the diverse and most represented species.</li> </ul>
589 590 591 592 593	<ul> <li><i>Reports</i>, 5(4), 395–402. https://doi.org/10.3892/br.2016.747</li> <li>Ferrero, G., Cordero, F., Tarallo, S., Arigoni, M., Riccardo, F., Gallo, G. &amp; Naccarati, A. (2017). Small non-coding RNA profiling in human biofluids and surrogate tissues from healthy individuals: Description of the diverse and most represented species. <i>Oncotarget</i>, 9(3), 3097–3111. https://doi.org/10.18632/oncotarget.23203</li> </ul>
589 590 591 592 593 594	<ul> <li><i>Reports</i>, 5(4), 395–402. https://doi.org/10.3892/br.2016.747</li> <li>Ferrero, G., Cordero, F., Tarallo, S., Arigoni, M., Riccardo, F., Gallo, G. &amp; Naccarati, A. (2017). Small non-coding RNA profiling in human biofluids and surrogate tissues from healthy individuals: Description of the diverse and most represented species. <i>Oncotarget</i>, 9(3), 3097–3111. https://doi.org/10.18632/oncotarget.23203</li> <li>Fraser, G. E., Jaceldo-Siegl, K., Orlich, M., Mashchak, A., Sirirat, R. &amp; Knutsen, S. (2020).</li> </ul>
589 590 591 592 593 594 595	<ul> <li><i>Reports</i>, 5(4), 395–402. https://doi.org/10.3892/br.2016.747</li> <li>Ferrero, G., Cordero, F., Tarallo, S., Arigoni, M., Riccardo, F., Gallo, G. &amp; Naccarati, A. (2017). Small non-coding RNA profiling in human biofluids and surrogate tissues from healthy individuals: Description of the diverse and most represented species. <i>Oncotarget</i>, 9(3), 3097–3111. https://doi.org/10.18632/oncotarget.23203</li> <li>Fraser, G. E., Jaceldo-Siegl, K., Orlich, M., Mashchak, A., Sirirat, R. &amp; Knutsen, S. (2020). Dairy, soy, and risk of breast cancer: Those confounded milks. <i>International Journal of</i></li> </ul>

- circulation. *Annual Review of Nutrition*, *36*, 301–336. https://doi.org/10.1146/annurev nutr-071715-050711
- Fromm, B., Tosar, J. P., Lu, Y., Halushka, M. K. & Witwer, K. W. (2018). Human and cow
- have identical miR-21-5p and miR-30a-5p sequences, which are likely unsuited to study
- dietary uptake from cow milk. *Journal of Nutrition*, *148*(9), 1506–1507.
- 604 https://doi.org/10.1093/jn/nxy144
- Galley, J. D. & Besner, G. E. (2020). The therapeutic potential of breast milk-derived
  extracellular vesicles. *Nutrients*, *12*(3):745. https://doi.org/10.3390/nu12030745
- 607 Gessner, I., Yu, X., Jüngst, C., Klimpel, A., Wang, L., Fischer, T. & Mathur, S. (2019).
- 608 Selective capture and purification of microRNA and intracellular proteins through
- antisense-vectorized magnetic nanobeads. *Scientific Reports*, *9*(1), 2069.
- 610 https://doi.org/10.1038/s41598-019-39575-7
- 611 Golan-Gerstl, R., Elbaum Shiff, Y., Moshayoff, V., Schecter, D., Leshkowitz, D. & Reif, S.
- 612 (2017). Characterization and biological function of milk-derived miRNA. *Molecular*
- 613 *Nutrition & Food Research*, *61*(10), 1700009. https://doi.org/10.1002/mnfr.201700009
- 614 Gorji-Bahri, G., Hashemi, A. & Moghimi, H. R. (2018). ExomiRs: A novel strategy in cancer
- 615 diagnosis and therapy. *Current Gene Therapy*, *18*(6), 336–350.
- 616 https://doi.org/10.2174/1566523218666181017163204
- 617 Gupta, S. K., Bang, C. & Thum, T. (2010). Circulating microRNA as biomarkers and
- 618 potential paracrine mediators of cardiovascular disease. *Circulation. Cardiovascular*
- 619 *Genetics*, *3*(5), 484–488. https://doi.org/10.1161/CIRCGENETICS.110.958363
- Ha, M., Pang, M., Agarwal, V. & Chen, Z. J. (2008). Interspecies regulation of microRNA
- and their targets. *Biochimica et Biophysica Acta*, 1779(11), 735–742.

622 https://doi.org/10.1016/j.bbagrm.2008.03.004

- Heintz-Buschart, A., Yusuf, D., Kaysen, A., Etheridge, A., Fritz, J. V, May, P. & Wilmes, P.
  (2018). Small RNA profiling of low biomass samples: Identification and removal of
  contaminants. *BMC Biology*, *16*(1), 52. https://doi.org/10.1186/s12915-018-0522-7
- 626 Howard, K. M., Jati Kusuma, R., Baier, S. R., Friemel, T., Markham, L., Vanamala, J. &
- E27 Zempleni, J. (2015). Loss of miRNA during processing and storage of cow's (*Bos*

628 *taurus*) milk. *Journal of Agricultural and Food Chemistry*, 63(2), 588–592.

- 629 https://doi.org/10.1021/jf505526w
- Huang, H., Pham, Q., Davis, C. D., Yu, L. & Wang, T. T. Y. (2020). Delineating effect of

631 corn microRNA and matrix, ingested as whole food, on gut microbiota in a rodent
632 model. *Food Science & Nutrition*, n/a(n/a). https://doi.org/10.1002/fsn3.1672

Izumi, H, Kosaka, N., Shimizu, T., Sekine, K., Ochiya, T. & Takase, M. (2012). Bovine milk

634 contains microRNA and messenger RNA that are stable under degradative conditions.

- 635 *Journal of Dairy Science*, 95(9), 4831–4841. https://doi.org/10.3168/jds.2012-5489
- Izumi, Hirohisa, Tsuda, M., Sato, Y., Kosaka, N., Ochiya, T., Iwamoto, H. & Takeda, Y.
- 637 (2015). Bovine milk exosomes contain microRNA and mRNA and are taken up by

human macrophages. *Journal of Dairy Science*, *98*(5), 2920–2933.

- 639 https://doi.org/10.3168/jds.2014-9076
- 640 Jeyaraman, M. M., Abou-Setta, A. M., Grant, L., Farshidfar, F., Copstein, L., Lys, J. &
- 641 Zarychanski, R. (2019). Dairy product consumption and development of cancer: An
- 642 overview of reviews. *BMJ Open*, 9(1), e023625. https://doi.org/10.1136/bmjopen-2018-
- 643 023625
- 644 Jin, H. Y., Gonzalez-Martin, A., Miletic, A. V, Lai, M., Knight, S., Sabouri-Ghomi, M. &

645	Xiao, C. (2015). Transfection of microRNA mimics should be used with caution.
646	Frontiers in Genetics, 6, 340. https://doi.org/10.3389/fgene.2015.00340
647	Jin, W., Ibeagha-Awemu, E. M., Liang, G., Beaudoin, F., Zhao, X. & Guan, L. L. (2014).
648	Transcriptome microRNA profiling of bovine mammary epithelial cells challenged with
649	Escherichia coli or Staphylococcus aureusbacteria reveals pathogen directed microRNA
650	expression profiles. BMC Genomics, 15(1), 181. https://doi.org/10.1186/1471-2164-15-
651	181
652	Jin, Y., Wong, Y. S., Goh, B. K. P., Chan, C. Y., Cheow, P. C., Chow, P. K. H. & Lee, C. G.
653	L. (2019). Circulating microRNA as potential diagnostic and prognostic biomarkers in
654	hepatocellular carcinoma. Scientific Reports, 9(1), 10464.
655	https://doi.org/10.1038/s41598-019-46872-8
656	Jodar, M. & Anton, E. (2018). Small RNA present in semen and their role in reproduction.
657	Reproductomics, 109-123. https://doi.org/10.1016/B978-0-12-812571-7.00008-3
658	Kang, W., Bang-Berthelsen, C. H., Holm, A., Houben, A. J. S., Müller, A. H., Thymann, T.
659	& Friedländer, M. R. (2017). Survey of 800+ data sets from human tissue and body fluid
660	reveals xenomiRs are likely artifacts. RNA, 23(4), 433-445.
661	https://doi.org/10.1261/rna.059725.116
662	Kelleher, S. L., Gagnon, A., Rivera, O. C., Hicks, S. D., Carney, M. C. & Alam, S. (2019).
663	Milk-derived miRNA profiles elucidate molecular pathways that underlie breast
664	dysfunction in women with common genetic variants in SLC30A2. Scientific Reports,
665	9(1), 12686. https://doi.org/10.1038/s41598-019-48987-4
666	Kincaid, R. P. & Sullivan, C. S. (2012). Virus-encoded microRNA: An overview and a look
667	to the future. PLoS Pathogens, 8(12), e1003018.
668	https://doi.org/10.1371/journal.ppat.1003018
	28

- 669 Kirchner, B., Buschmann, D., Paul, V. & Pfaffl, M. W. (2020). Postprandial transfer of
- 670 colostral extracellular vesicles and their protein and miRNA cargo in neonatal calves.

671 *PloS One*, *15*(2), e0229606. https://doi.org/10.1371/journal.pone.0229606

- Kosaka, N., Izumi, H., Sekine, K. & Ochiya, T. (2010). microRNA as a new immune-
- 673 regulatory agent in breast milk. *Silence*, *1*(1), 7. https://doi.org/10.1186/1758-907X-1-7
- Kumarswamy, R., Volkmann, I. & Thum, T. (2011). Regulation and function of miRNA-21
  in health and disease. *RNA Biology*, 8(5), 706–713.
- 676 https://doi.org/10.4161/rna.8.5.16154
- 677 Lässer, C., Alikhani, V. S., Ekström, K., Eldh, M., Paredes, P. T., Bossios, A. & Valadi, H.

678 (2011). Human saliva, plasma and breast milk exosomes contain RNA: Uptake by

- 679 macrophages. *Journal of Translational Medicine*, *9*, 9. https://doi.org/10.1186/1479 680 5876-9-9
- Le Doare, K., Holder, B., Bassett, A. & Pannaraj, P. S. (2018). Mother's milk: A purposeful
- 682 contribution to the development of the infant microbiota and immunity. *Frontiers in*
- 683 *Immunology*, 9, 361. https://doi.org/10.3389/fimmu.2018.00361
- Le Huërou-Luron, I., Blat, S. & Boudry, G. (2010). Breast- v. formula-feeding: Impacts on
- the digestive tract and immediate and long-term health effects. *Nutrition Research*

686 *Reviews*, 23(1), 23–36. https://doi.org/10.1017/S0954422410000065

- 687 Ledda, B., Ottaggio, L., Izzotti, A., Sukkar, S. G. & Miele, M. (2020). Small RNA in
- eucaryotes: New clues for amplifying microRNA benefits. *Cell & Bioscience*, 10(1), 1.
  https://doi.org/10.1186/s13578-019-0370-3
- 690 Lee, H.J. (2019). Microbe-host communication by small RNA in extracellular vesicles:
- 691 Vehicles for transkingdom RNA Transportation. *International Journal of Molecular*

692

*Sciences*, 20(6). https://doi.org/10.3390/ijms20061487

- 693 Lekka, E. & Hall, J. (2018). Noncoding RNA in disease. *FEBS Letters*, 592(17), 2884–2900.
  694 https://doi.org/10.1002/1873-3468.13182
- Li, R., Beaudoin, F., Ammah, A. A., Bissonnette, N., Benchaar, C., Zhao, X., Ibeagha-
- Awemu, E. M. (2015). Deep sequencing shows microRNA involvement in bovine
- 697 mammary gland adaptation to diets supplemented with linseed oil or safflower oil. *BMC*
- 698 *Genomics*, *16*, 884. https://doi.org/10.1186/s12864-015-1965-7
- Li, Y., Deng, X., Zeng, X. & Peng, X. (2016). The role of mir-148a in cancer. Journal of
- 700 *Cancer*, 7(10), 1233–1241. https://doi.org/10.7150/jca.14616
- Li, Z., Xu, R. & Li, N. (2018). MicroRNA from plants to animals, do they define a new
- messenger for communication? *Nutrition & Metabolism*, *15*(1), 68.
- 703 https://doi.org/10.1186/s12986-018-0305-8
- Liang, G., Zhu, Y., Sun, B., Shao, Y., Jing, A., Wang, J. & Xiao, Z. (2014). Assessing the
- survival of exogenous plant microRNA in mice. *Food Science & Nutrition*, 2(4), 380–
  388. https://doi.org/10.1002/fsn3.113
- Liang, H., Huang, L., Cao, J., Zen, K., Chen, X. & Zhang, C.-Y. (2012). Regulation of
- mammalian gene expression by exogenous microRNA. *Wiley Interdisciplinary Reviews*.
- 709 *RNA*, *3*(5), 733–742. https://doi.org/10.1002/wrna.1127
- Liang, H., Zhang, S., Fu, Z., Wang, Y., Wang, N., Liu, Y. & Zhang, C.Y. (2015). Effective
- 711 detection and quantification of dietetically absorbed plant microRNA in human plasma.
- *The Journal of Nutritional Biochemistry*, *26*(5), 505–512.
- 713 https://doi.org/10.1016/j.jnutbio.2014.12.002
- Liao, Y., Du, X., Li, J. & Lönnerdal, B. (2017). Human milk exosomes and their microRNA

- survive digestion *in vitro* and are taken up by human intestinal cells. *Molecular*
- 716 *Nutrition & Food Research*, *61*(11), e1701050. https://doi.org/10.1002/mnfr.201700082
- 717 Link, J., Thon, C., Schanze, D., Steponaitiene, R., Kupcinskas, J., Zenker, M. & Link, A.
- 718 (2019). Food-derived xeno-microRNA: Influence of diet and detectability in
- 719 gastrointestinal tract-proof-of-principle study. *Molecular Nutrition & Food Research*,
- 720 *63*(2), e1800076. https://doi.org/10.1002/mnfr.201800076
- Liu, H., Lei, C., He, Q., Pan, Z., Xiao, D. & Tao, Y. (2018). Nuclear functions of mammalian
- 722 MicroRNA in gene regulation, immunity and cancer. *Molecular Cancer*, 17(1), 64.
- 723 https://doi.org/10.1186/s12943-018-0765-5
- 724 Liu, S., da Cunha, A. P., Rezende, R. M., Cialic, R., Wei, Z., Bry, L. & Weiner, H. L. (2016).
- The host shapes the gut microbiota via fecal microRNA. *Cell Host & Microbe*, 19(1),
- 726 32–43. https://doi.org/10.1016/j.chom.2015.12.005
- Lu, W., Chen, H., Niu, Y., Wu, H., Xia, D. & Wu, Y. (2016). Dairy products intake and
- cancer mortality risk: A meta-analysis of 11 population-based cohort studies. *Nutrition*
- 729 *Journal*, 15(1), 91. https://doi.org/10.1186/s12937-016-0210-9
- 730 Lukasik, A., Brzozowska, I., Zielenkiewicz, U. & Zielenkiewicz, P. (2017). Detection of
- 731 plant miRNA abundance in human breast milk. *International Journal of Molecular*
- 732 *Sciences*, *19*(1). https://doi.org/10.3390/ijms19010037
- 733 Ma, S., Tong, C., Ibeagha-Awemu, E. M. & Zhao, X. (2019). Identification and
- characterization of differentially expressed exosomal microRNA in bovine milk infected
- with Staphylococcus aureus. *BMC Genomics*, 20(1), 934.
- 736 https://doi.org/10.1186/s12864-019-6338-1
- 737 Manca, S., Upadhyaya, B., Mutai, E., Desaulniers, A. T., Cederberg, R. A., White, B. R. &

- 738 Zempleni, J. (2018). Milk exosomes are bioavailable and distinct microRNA cargos
- have unique tissue distribution patterns. *Scientific Reports*, 8(1), 11321.
- 740 https://doi.org/10.1038/s41598-018-29780-1
- 741 Martin, C. R., Ling, P.R. & Blackburn, G. L. (2016). Review of infant feeding: Key features
- of breast milk and infant formula. *Nutrients*, 8(5), 279.
- 743 https://doi.org/10.3390/nu8050279
- Martin, M. (2017). Michael L. Power and Jay Schulkin: Milk: The biology of lactation.
- 745 International Journal of Primatology, 38(1), 100–103. https://doi.org/10.1007/s10764-
- 746 017-9950-4
- 747 Melnik, B. C. (2017). Milk disrupts p53 and DNMT1, the guardians of the genome:
- 748Implications for acne vulgaris and prostate cancer. Nutrition & Metabolism, 14(1), 55.
- 749 https://doi.org/10.1186/s12986-017-0212-4
- 750 Melnik, B. C. & Schmitz, G. (2017). MicroRNA: Milk's epigenetic regulators. Best Practice
- 751 & Research. Clinical Endocrinology & Metabolism, 31(4), 427–442.
- 752 https://doi.org/10.1016/j.beem.2017.10.003
- 753 Melnik, B. C. & Schmitz, G. (2019). Exosomes of pasteurized milk:: Potential pathogens of
- 754 Western diseases. *Journal of Translational Medicine*, *17*(1), 3.
- 755 https://doi.org/10.1186/s12967-018-1760-8
- 756 Mi, S., Zhang, J., Zhang, W. & Huang, R. S. (2013). Circulating microRNA as biomarkers
- for inflammatory diseases. *MicroRNA*, 2(1), 63–71.
- 758 https://doi.org/10.2174/2211536611302010007
- 759 Mirza, A. H., Kaur, S., Nielsen, L. B., Størling, J., Yarani, R., Roursgaard, M. & Pociot, F.
- 760 (2019). Breast milk-derived extracellular vesicles enriched in exosomes from mothers

- 761 with type 1 diabetes contain aberrant levels of microRNA. *Frontiers in Immunology*,
- 762 *10*, 2543. https://doi.org/10.3389/fimmu.2019.02543
- 763 Muroya, S., Hagi, T., Kimura, A., Aso, H., Matsuzaki, M. & Nomura, M. (2016). Lactogenic
- hormones alter cellular and extracellular microRNA expression in bovine mammary
- repithelial cell culture. *Journal of Animal Science and Biotechnology*, 7, 8.
- 766 https://doi.org/10.1186/s40104-016-0068-x
- Nguyen, T. (2020, June). Unravelling the mysteries of microRNA in breast milk. *Nature*.
  England. https://doi.org/10.1038/d41586-020-01768-w
- 769 Oh, S., Park, M.R., Ryu, S., Maburutse, B., Kim, J.-U. & Kim, Y. (2017). Quantitative
- analysis of milk-derived microRNA and microbiota during the manufacturing and
- ripening of soft cheese. *Journal of Microbiology and Biotechnology*, 27(9), 1566–1575.
- 772 https://doi.org/10.4014/jmb.1705.05006
- Park, N. J., Zhou, H., Elashoff, D., Henson, B. S., Kastratovic, D. A., Abemayor, E. & Wong,
- D. T. (2009). Salivary microRNA: Discovery, characterization, and clinical utility for
- oral cancer detection. *Clinical Cancer Research* An Official Journal of the American
- 776 Association for Cancer Research, 15(17), 5473–5477. https://doi.org/10.1158/1078-
- 777 0432.CCR-09-0736
- Pathan, M., Fonseka, P., Chitti, S. V, Kang, T., Sanwlani, R., Van Deun, J. & Mathivanan, S.
- (2019). Vesiclepedia 2019: A compendium of RNA, proteins, lipids and metabolites in
  extracellular vesicles. *Nucleic Acids Research*, 47(D1), D516–D519.
- 781 https://doi.org/10.1093/nar/gky1029
- 782 Perge, P., Nagy, Z., Decmann, A., Igaz, I. & Igaz, P. (2017). Potential relevance of
- 783 microRNA in inter-species epigenetic communication, and implications for disease
- 784 pathogenesis. *RNA Biology*, *14*(4), 391–401.

785

https://doi.org/10.1080/15476286.2016.1251001

- 786 Pieters, B. C. H., Arntz, O. J., Bennink, M. B., Broeren, M. G. A., van Caam, A. P. M.,
- 787 Koenders, M. I., van Lent, P. L., van den Berg, W. B., de Vries, M., van der Kraan, P.
- 788 M., van de Loo, F. A. J. (2015). Commercial cow milk contains physically stable
- extracellular vesicles expressing immunoregulatory TGF-β. *Plos*  $\frac{10}{3}$ , *10*(3), e0121123.
- 790 https://doi.org/10.1371/journal.pone.0121123
- 791 Pirim, D. & Dogan, B. (2020). In silico identification of putative roles of food-derived xeno-
- mirs on diet-associated cancer. *Nutrition and Cancer*, 72(3), 481–488.
- 793 https://doi.org/10.1080/01635581.2019.1670854
- 794 Quan, S.Y., Nan, X.M., Wang, K., Zhao, Y.G., Jiang, L.S., Yao, J. H. & Xiong, B.H. (2020).
- 795 Replacement of forage fiber with non-forage fiber sources in dairy cow diets changes
- milk extracellular vesicle-miRNA expression. *Food & Function*, 11(3), 2154–2162.
- 797 https://doi.org/10.1039/c9fo03097b
- 798 Quintanilha, B. J., Reis, B. Z., Duarte, G. B. S., Cozzolino, S. M. F. & Rogero, M. M. (2017).
- 799 Nutrimiromics: Role of microRNA and nutrition in modulating inflammation and
- 800 chronic diseases. *Nutrients*, 9(11), 1168. https://doi.org/10.3390/nu9111168
- 801 Rani, P., Vashisht, M., Golla, N., Shandilya, S., Onteru, S. K. & Singh, D. (2017). Milk
- 802 miRNA encapsulated in exosomes are stable to human digestion and permeable to
- 803 intestinal barrier *in vitro*. *Journal of Functional Foods*, *34*, 431–439.
- 804 https://doi.org/https://doi.org/10.1016/j.jff.2017.05.009
- Rani, P., Yenuganti, V. R., Shandilya, S., Onteru, S. K. & Singh, D. (2017). miRNA: The
- hidden bioactive component of milk. *Trends in Food Science & Technology*, 65, 94–
- 807 102. https://doi.org/https://doi.org/10.1016/j.tifs.2017.05.007

34

- 808 Rebane, A. & Akdis, C. A. (2013). MicroRNA: Essential players in the regulation of
- inflammation. *The Journal of Allergy and Clinical Immunology*, *132*(1), 15–26.
- 810 https://doi.org/10.1016/j.jaci.2013.04.011
- 811 Redell, J. B., Moore, A. N., Ward, N. H., Hergenroeder, G. W. & Dash, P. K. (2010). Human
- traumatic brain injury alters plasma microRNA levels. *Journal of Neurotrauma*, 27(12),
- 813 2147–2156. https://doi.org/10.1089/neu.2010.1481
- 814 Roth, C., Rack, B., Müller, V., Janni, W., Pantel, K. & Schwarzenbach, H. (2010).
- 815 Circulating microRNA as blood-based markers for patients with primary and metastatic
- breast cancer. Breast Cancer Research, 12(6), R90. https://doi.org/10.1186/bcr2766
- 817 Sanchita, Trivedi, R., Asif, M. H. & Trivedi, P. K. (2018). Dietary plant miRNA as an
- 818 augmented therapy: Cross-kingdom gene regulation. *RNA Biology*, *15*(12), 1433–1439.
  819 https://doi.org/10.1080/15476286.2018.1551693
- 820 Santamaria-Martos, F., Benítez, I., Ortega, F., Zapater, A., Giron, C., Pinilla, L., Pascual, L.,
- 821 Cortijo, A., Dalmases, M., Fernandez-Real, J.M., Barbé, F. & Sánchez-de-la-Torre, M.
- 822 (2019). Circulating microRNA profile as a potential biomarker for obstructive sleep
- apnea diagnosis. *Scientific Reports*, 9(1), 13456. https://doi.org/10.1038/s41598-01949940-1
- 825 Sarshad, A. A., Juan, A. H., Muler, A. I. C., Anastasakis, D. G., Wang, X., Genzor, P., Feng,
- 826 X., Tsai, P. F., Sun, H. W., Haase, A. D., Sartorelli, V. & Hafner, M. (2018). Argonaute-
- miRNA complexes silence target mRNA in the nucleus of mammalian stem cells.
- 828 *Molecular Cell*, 71(6), 1040-1050.e8. https://doi.org/10.1016/j.molcel.2018.07.020
- 829 Shmaryahu, A., Carrasco, M. & Valenzuela, P. D. T. (2014). Prediction of bacterial
- 830 microRNA and possible targets in human cell transcriptome. *Journal of Microbiology*
- 831 (Seoul, Korea), 52(6), 482–489. https://doi.org/10.1007/s12275-014-3658-3

- 832 Simpson, M. R., Brede, G., Johansen, J., Johnsen, R., Storrø, O., Sætrom, P. & Øien, T.
- 833 (2015). Human breast milk miRNA, maternal probiotic supplementation and atopic
- dermatitis in offspring. *PloS One*, *10*(12), e0143496.
- 835 https://doi.org/10.1371/journal.pone.0143496
- 836 Snow, J. W., Hale, A. E., Isaacs, S. K., Baggish, A. L. & Chan, S. Y. (2013). Ineffective
- delivery of diet-derived microRNA to recipient animal organisms. RNA Biology, 10(7),
- 838 1107–1116. https://doi.org/10.4161/rna.24909
- 839 Stephen, B. J., Pareek, N., Saeed, M., Kausar, M. A., Rahman, S. & Datta, M. (2020). Xeno-
- 840 mirna in maternal-infant immune crosstalk: An aid to disease alleviation. Frontiers in
- 841 *Immunology*, *11*, 404. https://doi.org/10.3389/fimmu.2020.00404
- 842 Sun, J., Aswath, K., Schroeder, S. G., Lippolis, J. D., Reinhardt, T. A. & Sonstegard, T. S.
- 843 (2015). MicroRNA expression profiles of bovine milk exosomes in response to
  844 Staphylococcus aureus infection. *BMC Genomics*, *16*(1), 806.
- 845 https://doi.org/10.1186/s12864-015-2044-9
- 846 Svoronos, A. A., Engelman, D. M. & Slack, F. J. (2016). OncomiR or tumor suppressor? The
- duplicity of microRNA in cancer. *Cancer Research*, 76(13), 3666–3670.
- 848 https://doi.org/10.1158/0008-5472.CAN-16-0359
- 849 Teng, Y., Ren, Y., Sayed, M., Hu, X., Lei, C., Kumar, A. & Zhang, H.-G. (2018). Plant-
- derived exosomal microRNA shape the gut microbiota. *Cell Host & Microbe*, 24(5),
  637-652.e8. https://doi.org/10.1016/j.chom.2018.10.001
- 852 Testroet, E. D., Shome, S., Testroet, A., Reecy, J., Jernigan, R. L., Zhu, M. & Beitz, D.
- 853 (2018). Profiling of the exosomal cargo of bovine milk reveals the presence of immune-
- and growth-modulatory ncRNA. *Iowa State University Animal Industry Report*, 15(1).
- doi: https://doi.org/10.31274/ans\_air-180814-330

- Title, A. C., Denzler, R. & Stoffel, M. (2015). Uptake and function studies of maternal milk-
- derived microRNA. *The Journal of Biological Chemistry*, 290(39), 23680–23691.

858 https://doi.org/10.1074/jbc.M115.676734

- Title, A., Denzler, R. & Stoffel, M. (2015). Reply to Diet-responsive MicroRNA Are Likely
  Exogenous. *The Journal of Biological Chemistry*. 290, 25198.
- 861 https://doi.org/10.1074/jbc.L115.688358
- 862 Tomé-Carneiro, J., Fernández-Alonso, N., Tomás-Zapico, C., Visioli, F., Iglesias-Gutierrez,
- E. & Dávalos, A. (2018). Breast milk microRNA harsh journey towards potential effects
- in infant development and maturation. Lipid encapsulation can help. *Pharmacological*
- 865 *Research*, *132*, 21–32. https://doi.org/10.1016/j.phrs.2018.04.003
- 866 Tong, L., Hao, H., Zhang, X., Zhang, Z., Lv, Y., Zhang, L. & Yi, H. (2020). Oral
- administration of bovine milk-derived extracellular vesicles alters the gut microbiota and
- 868 enhances intestinal immunity in mice. *Molecular Nutrition & Food Research*, 64(8),
- e1901251. https://doi.org/10.1002/mnfr.201901251
- 870 Turchinovich, A, Samatov, T. R., Tonevitsky, A. G. & Burwinkel, B. (2013). Circulating
- miRNA: Cell-cell communication function? *Frontiers in Genetics*, *4*, 119.
- 872 https://doi.org/10.3389/fgene.2013.00119
- Turchinovich, Andrey, Weiz, L., Langheinz, A. & Burwinkel, B. (2011). Characterization of
- extracellular circulating microRNA. *Nucleic Acids Research*, *39*(16), 7223–7233.
- 875 https://doi.org/10.1093/nar/gkr254
- Tzifi, F., Grammeniatis, V. & Papadopoulos, M. (2014). Soy- and rice-based formula and
- 877 infant allergic to cow's milk. *Endocrine, Metabolic & Immune Disorders Drug Targets*,
- 878 *14*(1), 38–46. https://doi.org/10.2174/1871530314666140121144604

879	van Herwijnen, M. J. C., Driedonks, T. A. P., Snoek, B. L., Kroon, A. M. T., Kleinjan, M.,
880	Jorritsma, R. & Wauben, M. H. M. (2018). Abundantly present miRNA in milk-derived
881	extracellular vesicles are conserved between mammals. Frontiers in Nutrition, 5, 81.
882	https://doi.org/10.3389/fnut.2018.00081
883	Van Hese, I., Goossens, K., Vandaele, L. & Opsomer, G. (2020). Invited review: MicroRNA
884	in bovine colostrum-Focus on their origin and potential health benefits for the calf.
885	Journal of Dairy Science, 103(1), 1-15. https://doi.org/10.3168/jds.2019-16959
886	Vaucheret, H. & Chupeau, Y. (2012). Ingested plant miRNA regulate gene expression in
887	animals. Cell Research, 22(1), 3-5. https://doi.org/10.1038/cr.2011.164
888	Wang, D., Liang, G., Wang, B., Sun, H., Liu, J. & Guan, L. L. (2016). Systematic
889	microRNAome profiling reveals the roles of microRNA in milk protein metabolism and
890	quality: Insights on low-quality forage utilization. Scientific Reports, 6, 21194.
891	https://doi.org/10.1038/srep21194
892	Wang, L., Sadri, M., Giraud, D. & Zempleni, J. (2018). RNAe H2-dependent polymerase
893	chain reaction and elimination of confounders in sample collection, storage, and analysis
894	strengthen evidence that microRNA in bovine milk are bioavailable in humans. Journal
895	of Nutrition, 148(1), 153-159. https://doi.org/10.1093/jn/nxx024
896	Weber, J. A., Baxter, D. H., Zhang, S., Huang, D. Y., Huang, K. H., Lee, M. J., Galas, D. J.
897	& Wang, K. (2010). The microRNA spectrum in 12 body fluids. Clinical Chemistry,
898	56(11), 1733–1741. https://doi.org/10.1373/clinchem.2010.147405
899	Witwer, K. W. (2014). Diet-responsive mammalian miRNA are likely endogenous. Journal
900	of Nutrition. 144, 1880-1881. https://doi.org/10.3945/jn.114.202523

901 Witwer, K. W. (2018). Alternative miRNA? Human sequences misidentified as plant miRNA

- 902 in plant studies and in human plasma. *F1000Research*, 7, 244.
- 903 https://doi.org/10.12688/f1000research.14060.1
- 904 Witwer, K. W., McAlexander, M. A., Queen, S. E. & Adams, R. J. (2013). Real-time
- 905 quantitative PCR and droplet digital PCR for plant miRNA in mammalian blood provide
- 906 little evidence for general uptake of dietary miRNA: Limited evidence for general
- 907 uptake of dietary plant xenomiRs. *RNA Biology*, *10*(7), 1080–1086.
- 908 https://doi.org/10.4161/rna.25246
- 909 Witwer, K. W. & Zhang, C.Y. (2017). Diet-derived microRNA: Unicorn or silver bullet?
- 910 *Genes & Nutrition*, 12(1), 15. https://doi.org/10.1186/s12263-017-0564-4
- 911 Wolf, T., Baier, S. R. & Zempleni, J. (2015). The intestinal transport of bovine milk
- 912 exosomes is mediated by endocytosis in human colon carcinoma caco-2 cells and rat
- small intestinal IEC-6 cells. *Journal of Nutrition*, *145*(10), 2201–2206.
- 914 https://doi.org/10.3945/jn.115.218586
- 915 Wu, D., Kittana, H., Shu, J., Kachman, S. D., Cui, J., Ramer-Tait, A. E. & Zempleni, J.
- 916 (2019). Dietary depletion of milk exosomes and their microRNA cargos elicits a
- 917 depletion of miR-200a-3p and elevated intestinal inflammation and chemokine (C-X-C
- 918 motif) ligand 9 expression in mdr1a-/- mice. *Current Developments in Nutrition*, 3(12).
- 919 https://doi.org/10.1093/cdn/nzz122
- 920 Yang, J., Elbaz-Younes, I., Primo, C., Murungi, D. & Hirschi, K. D. (2018). Intestinal
- 921 permeability, digestive stability and oral bioavailability of dietary small RNA. *Scientific*
- 922 *Reports*, 8(1), 10253. https://doi.org/10.1038/s41598-018-28207-1
- 923 Yang, J., Farmer, L. M., Agyekum, A. A. A., Elbaz-Younes, I. & Hirschi, K. D. (2015).
- 924 Detection of an abundant plant-based small rna in healthy consumers. *PloS One*, *10*(9),
- 925 e0137516. https://doi.org/10.1371/journal.pone.0137516

926	Yang, J., Farmer, L. M., Agyekum, A. A. A. & Hirschi, K. D. (2015). Detection of dietary
927	plant-based small RNA in animals. Cell Research, 25(4), 517–520.
928	https://doi.org/10.1038/cr.2015.26

929 Yang, J., Hirschi, K. D. & Farmer, L. M. (2015). Dietary RNA: New stories regarding oral

930 delivery. *Nutrients*, 7(5), 3184–3199. https://doi.org/10.3390/nu7053184

- 931 Yeri, A., Courtright, A., Reiman, R., Carlson, E., Beecroft, T., Janss, A., Van Keuren-Jensen,
- 932 K. (2017). Total extracellular small RNA profiles from plasma, saliva, and urine of
- healthy subjects. *Scientific Reports*, 7, 44061. https://doi.org/10.1038/srep44061
- 934 Yu, S., Zhao, Z., Sun, L. & Li, P. (2017). Fermentation results in quantitative changes in
- 935 milk-derived exosomes and different effects on cell growth and survival. *Journal of*
- 936 *Agricultural and Food Chemistry*, 65(6), 1220–1228.
- 937 https://doi.org/10.1021/acs.jafc.6b05002
- 238 Zempleni, J., Aguilar-Lozano, A., Sadri, M., Sukreet, S., Manca, S., Wu, D. & Mutai, E.
- 939 (2016). Biological activities of extracellular vesicles and their cargos from bovine and
- human milk in humans and implications for infants. *Journal of Nutrition*, *147*(1), 3–10.
- 941 https://doi.org/10.3945/jn.116.238949
- Zempleni, J., Baier, S. R. & Hirschi, K. (2015). Diet-responsive microRNA are likely
  exogenous. *Journal of Biological Chemistry*, 290(41), 25197.
- 944 https://doi.org/10.1074/jbc.L115.687830
- 245 Zempleni, J., Baier, S. R., Howard, K. M. & Cui, J. (2015). Gene regulation by dietary
- 946 microRNA. *Canadian Journal of Physiology and Pharmacology*, *93*(12), 1097–1102.
- 947 https://doi.org/10.1139/cjpp-2014-0392
- 248 Zeng, J., Gupta, V. K., Jiang, Y., Yang, B., Gong, L. & Zhu, H. (2019). Cross-kingdom small

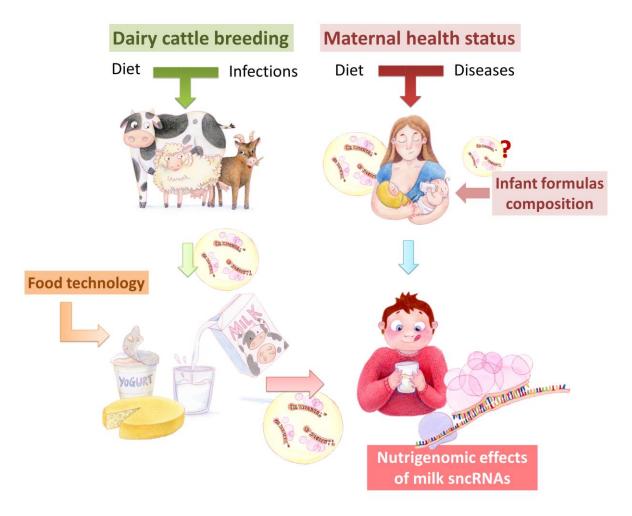
- 949 RNA among animals, plants and microbes. *Cells*, 8(4), 371.
- 950 https://doi.org/10.3390/cells8040371
- 251 Zhang, L., Chen, T., Yin, Y., Zhang, C.-Y. & Zhang, Y. L. (2019). Dietary microRNA-A
- 952 novel functional component of food. *Advances in Nutrition*, *10*(4), 711–721.
- 953 https://doi.org/10.1093/advances/nmy127
- 254 Zhang, L., Hou, D., Chen, X., Li, D., Zhu, L., Zhang, Y., Zhang, C.Y. (2012). Exogenous
- 955 plant MIR168a specifically targets mammalian LDLRAP1: Evidence of cross-kingdom
- regulation by microRNA. *Cell Research*, 22(1), 107–126.
- 957 https://doi.org/10.1038/cr.2011.158
- 958 Zhao, Q., Liu, Y., Zhang, N., Hu, M., Zhang, H., Joshi, T. & Xu, D. (2018). Evidence for
- 959 plant-derived xenomiRs based on a large-scale analysis of public small RNA
- sequencing data from human samples. *PloS One*, *13*(6), e0187519.
- 961 https://doi.org/10.1371/journal.pone.0187519
- 962 Zhao, Y., Cong, L. & Lukiw, W. J. (2018). Plant and animal microRNA (miRNA) and their
- 963 potential for inter-kingdom communication. *Cellular and Molecular Neurobiology*,
- 964 *38*(1), 133–140. https://doi.org/10.1007/s10571-017-0547-4
- 265 Zhou, F., Paz, H. A., Sadri, M., Fernando, S. C. & Zempleni, J. (2019). A diet defined by its
- content of bovine milk exosomes alters the composition of the intestinal microbiome in
- 967 C57BL/6 mice. *Am J Physiol Gastrointest Liver Physiol*, 317(5), G618-G624. doi:
- 968 10.1152/ajpgi.00160.2019.
- 269 Zhou, Q., Li, M., Wang, X., Li, Q., Wang, T., Zhu, Q. & Li, X. (2012). Immune-related
- 970 microRNA are abundant in breast milk exosomes. *International Journal of Biological*
- 971 Sciences, 8(1), 118–123. https://doi.org/10.7150/ijbs.8.118

#### **Figure legends**

#### Figure 1. Exogenous sncRNA can be conveyed by milk and exert inter-species effects in humans. SncRNA are contained in milk produced by dairy cattle, whose rearing conditions can modulate the final profile. Different food technologies can modulate the presence of sncRNA in milk that is used in human nutrition. SncRNA are also present in maternal milk, while their content seems to be depleted in infant formulas. The exposure to different profiles of exogenous sncRNA could contribute to the heterogeneous nutrigenomic effects exerted by milk and derivatives, and consequentially impact human health. Current knowledge of possible nutrigenomic effects induced by exogenous sncRNA is represented in the bottom right of the figure (+ likely to occur; - unlikely to occur/limited evidence).

# 996 Figures

997 Figure 1.



998

Conflict of Interest

# **Competing interests**

The authors declare no competing interests

Laura Bordoni, Phd

Lauro Bordoni

### **Author Agreement Statement**

The authors declare that this manuscript is original, has not been published before and is not currently being considered for publication elsewhere.

The authors confirm that the manuscript has been read and approved by all named authors and that there are no other persons who satisfied the criteria for authorship but are not listed.

The authors further confirm that the order of authors listed in the manuscript has been approved by all of us.

The authors understand that the Corresponding Author is the sole contact for the Editorial process. He/she is responsible for communicating with the other authors about progress, submissions of revisions and final approval of proofs

Signed by all authors as follows:

Dr. Laura Bordoni

Louveo Bordeni

Prof. Rosita Gabbianelli

Ainte Jelihaelli