



# Therapeutic approaches to coronavirus infection according to “One Health” concept

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## ABSTRACT

*Coronaviridae* constantly infect human and animals causing respiratory, gastroenteric or systemic diseases. Over time, these viruses have shown a marked ability to mutate, jumping over the human-animal barrier, thus becoming from enzootic to zoonotic. In the last years, numerous therapeutic protocols have been developed, mainly for severe acute respiratory syndromes in humans. The aim of this review is to summarize drugs or other approaches used in coronavirus infections focusing on different roles of these molecules or bacterial products on viral adhesion and replication or in modulating the host's immune system. Within the “One Health” concept, the study of viral pathogenic role and possible therapeutic approaches in both humans and animals is essential to protect public health.

## 1. Introduction

Over the past 17 years, three global outbreaks in humans have occurred due to viruses belonging to the genus betacoronavirus: SARS-Coronavirus-1 (SARS-CoV-1) in 2003, MERS-Coronavirus (MERS-CoV) in 2012 and SARS-Coronavirus-2 (SARS-CoV-2) in 2019 (World Health Organization, 2020; CDCP, 2019). These coronaviruses cause respiratory and gastroenteric diseases in mammals similar to alpha- and delta-coronaviruses (Masters and Perlman, 2013). Both genera belong to *Coronaviridae*, a family of positive sense, single-stranded RNA viruses with envelope, with large protrusions in the surface (van Regenmortel et al., 2000). The genome of these viruses encodes for structural proteins such as nucleocapsid protein (N), membrane protein (M), envelope protein (E), spike protein (S), and in some coronaviruses the envelope-associated hemagglutinin-esterase protein (HE) (Li, 2016; Schoeman and Fielding, 2019), and for non-structural proteins (nsp) such as proteases (nsp3 and nsp5) and RdRp (nsp12) (Elfiky et al., 2017; Kilianski and Baker, 2014). CoV entry occurs either via the plasma membrane with the help of cell surface proteases such as TMPRSS2 or via endocytosis, through the splitting of S proteins into two functional subunits: the S1 (harboring the receptor-binding domain) that binds of the surface unit, and the S2 (containing the membrane fusion domains) that allows fusion of viral and cellular membranes (Hoffmann et al., 2020; Tripet et al., 2004). Defined as enzootic, for a long time, some of these viruses have crossed the animal-human barrier becoming zoonotic (Chan et al.,

2013b), due to their ability to mutate and recombine adapting that way to new host range (Li, 2016). It is widely recognized that animals were the spillover source for SARS-CoV-2 infection in men (Schmiege et al., 2020; Di Teodoro et al., 2020). To deepen the link among animal-men-SARS-COV-2, the susceptibility of many animals to the virus has been investigated and it has been shown that animals such as cats, ferrets and minks are susceptible to the infection, while chickens and pigs are not; interestingly, respiratory *ex-vivo* organ cultures of cattle and sheep further suggested that also these animal species may be susceptible to the infection (Di Teodoro et al., 2020). Much information is available in veterinary medicine on animal coronaviruses, so that such knowledge may be of help also in human medicine to better understand how the virus behaves and how to face it. For example, in birds, CoVs have been detected in a lot of different species such as domestic fowl, turkeys, penguins, pigeons, duck, etc. But also bats, some rodents, swine, ruminants, horses, dogs, cats, ferrets, wolves, red foxes and many other wild carnivores are known to host CoVs (Decaro and Lorusso, 2020), suggesting that both human and veterinary medicine may benefit from a “One health” approach.

Studies in animal models allowed to characterize the viral pathogenesis of human coronaviruses (SARS-CoV-1 and MERS-CoV) and to actuate new therapeutically approaches (Sutton and Subbarao, 2015). In general, two types of therapeutic strategies can be recognized: the first aiming directly to counteract the etiological agent, the second aiming to control the host immune-response.

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## 2. Therapeutical approaches against the virus entry and the RNA replication

### 2.1. Nucleotide and nucleoside analogue inhibitors

These molecules are chemically synthesized of pyrimidine and purines and administered as precursors (Pruijssers and Denison, 2019). Nucleotide and nucleoside inhibitors compete with nucleotide substrate to bind active site of polymerase; these analogues can induce mutations that impair the RNA structure and RNA-protein functions or can cause chain termination in replicating viral genomes incorporating foreign nucleotides (Gao et al., 2020; Pruijssers and Denison, 2019). Numerous molecules (Fig. 1), have been proposed for coronavirus infections, and their activity has been demonstrated primarily in human coronavirus such as: acyclic fleximer nucleoside analogues (Peters et al., 2015); 6-Azauridine (Pyrce et al., 2006); Gemcitabine hydrochloride (Dyall et al., 2014); BCX4430 (Warren et al., 2014);  $\beta$ -d-N4-hydroxycytidine (Barnard et al., 2004) and the 1- $\beta$ -d-ribofuranosyl-1, 2,4-triazole-3-carboxamide (ribavirin) (Chan et al., 2013a); Mizoribine and Ribavirin (Saijo et al., 2005). However, the evidence of efficacy is inconclusive. Recently, the GS-441524 (parent nucleoside of Remdesivir), a broad-spectrum RNA polymerase inhibitor, has been proposed. It was developed in response to the Ebola outbreak (Mulangu et al., 2019), filoviruses, paramyxoviruses, and pneumoviruses (Lo et al., 2017). Its anti-CoV activity was shown in cats with feline infectious peritonitis (Murphy et al., 2018), in a mouse model of SARS-CoV-1 (Sheahan et al.,

2017) and in nonhuman primate model of MERS-CoV infection (Martinez, 2020), but its mechanism is still unclear. This adenosine nucleotide analogue can get incorporated into viral RNA and cause premature chain termination (Warren et al., 2016); by intracellular phosphorylation, the active NTP (Nucleoside Triphosphate) analog works as a competitor of the natural nucleoside triphosphates in RNA synthesis (Warren et al., 2016). Remdesivir can also interfere with the nsp12 polymerase, a non-structural protein which mediates the RNA replication (Shannon et al., 2020). Another nucleoside analog used for the treatment of feline infectious peritonitis in cats is Mutian X (Addie et al., 2020). This drug is a synthetic adenosine analogue associated with nicotinamide mononucleotide, Crocin I, S-Adenosylmethionine, Silymarin, thus stopping RNA replication (Addie et al., 2020). However, as an RNA virus, coronaviruses have an intrinsic genetic variability, which results in a high mutation rate and are therefore able to develop drug-resistance for these molecules (Agostini et al., 2018).

### 2.2. Viral protease inhibitors

By specific proteases, as papain-like protease and 3C-like protease (3CLpro), coronaviruses can cleave polyproteins and release non-structural proteins (nsp), as the NSP1–16, which have important functions in maturation or production of functional viral proteins (Sarma et al., 2020). These viral proteins, similar in human coronavirus and feline coronavirus, are key factors for RNA replication and transcription and, for this reason; they are target for protease inhibitors. A peptidyl

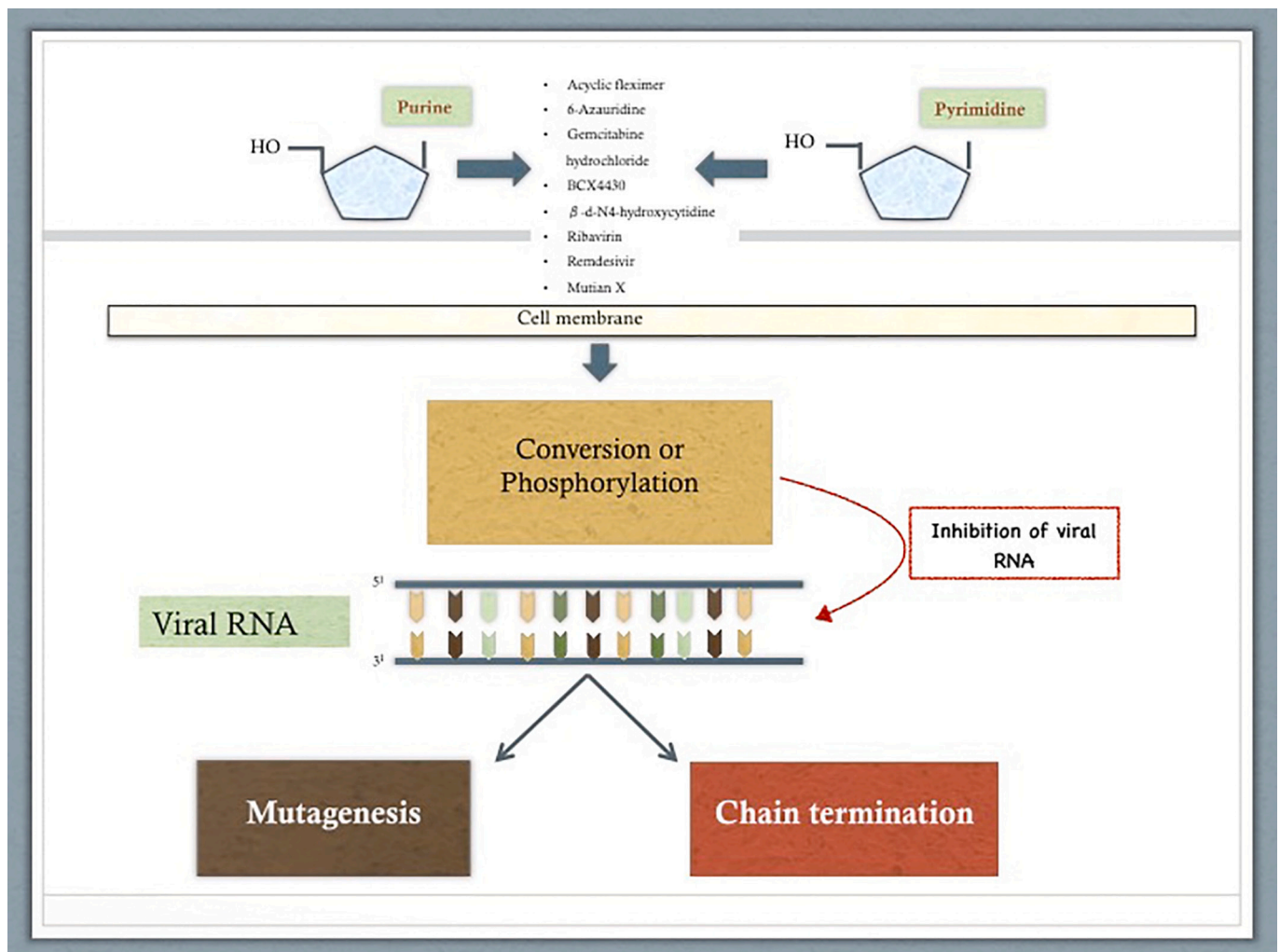


Fig. 1. Schematic representation of nucleotide and nucleoside inhibitors on viral RNA.

compounds targeting viral 3C-like protease has proven effective against feline infectious peritonitis in cat (Kim et al., 2015). This protease inhibitor, the GC376, has been tested in 20 sick cats, however the results did not demonstrate complete efficacy of the treatment (Pedersen et al., 2018). Anti-SARS-CoV-2 action has been demonstrated in protease inhibitors such as Lopinavir, Saquinavir (Dayer et al., 2017), and a formulation of two structurally related protease inhibitor for the treatment of SARS-CoV-2: Lopinavir associated with Ritonavir (Kaletra, Abbott Laboratories ® North Chicago) (Jin et al., 2020). In patients with HIV infection, it has been shown that Lopinavir can prevent subsequent infections of cells arresting maturation of HIV-1 (Cvetkovic and Goa, 2003). Lopinavir is an HIV-1 protease inhibitor, which hinders the formation of infectious virions but has poor bioavailability, therefore it comes co-formulated with ritonavir, which can inhibit the cytochrome P450 3A4 isoenzyme increasing lopinavir blood levels (Chandwani and Shuter, 2008). Finally, recent studies are demonstrating effectiveness of a triple combination composed by interferon beta-1b, Lopinavir–Ritonavir, and Ribavirin (nucleoside analogue) in patients with COVID-19, if administered within one week of symptom onset (Hung et al., 2020).

### 2.3. Cellular protease inhibitors

Viruses use specific proteins present in cell for entry (Böttcher et al., 2006; Glowacka et al., 2011). Through the cleavage of the surface glycoprotein spike (S) at two different sites, coronaviruses can enter in the host cell (Bestle et al., 2020). This step is mediated by host cell proteases. Among human and animal species these proteases are different and the species-specificity of coronaviruses is determined by the recognition of a functional receptor. Studies have shown that different proteases can activate the virus including trypsin, cathepsin, transmembrane serine proteases (in particular TMPRSS2), aminopeptidase N, and furin; some of which play major roles in a broad range of viruses (Heald-Sargent and Gallagher, 2012; Millet and Whittaker, 2015; Ou et al., 2020; Luan et al., 2020). In the cat, the receptor for type II feline coronavirus is the feline aminopeptidase N (FAPN) (Miguel et al., 2002). In the dog, Canine respiratory coronavirus (CRCoV) uses a caveolin-dependent endocytosis and a mechanism like that used by MERS-CoV by a transmembrane protease serine 2 (TMPRSS2) (Szczeplanski et al., 2018). The gene TMPRSS2 encodes proteins that belong to the serine protease family (Böttcher et al., 2006). In SARS-CoV-1 and MERS-CoV, the TMPRSS2 and the endosomal cysteine protease cathepsin L prime the S proteins, facilitating the entry of the virus and the splitting of the protein into two functional subunits: S1 which binds of the surface unit, and S2 which allows fusion of viral and cellular membranes (Hoffmann et al., 2020; Tripet et al., 2004). In general, the respiratory and gut epithelia are targeted for many coronavirus genera where the pathogen can cause a localized or systemic infection (Siddell et al., 2005). The metallopeptidase named angiotensin-converting enzyme 2 (ACE2) is a specific receptor present in different types of cells such as lung, gut, and prostate epithelial cells (Hamming et al., 2004; Song et al., 2020). In SARS-CoV-1, TMPRSS2 can modulate viral spread in the host by co-expression of ACE2 (Mossel et al., 2008), which binds to the S1 domain of the SARS-CoV-1 spike protein (Li et al., 2003). SARS-CoV-2 evolved to possess a furin cleavage site at its S1/S2 site essential to cleave viral fusion proteins allowing entry into human lung cells (Hoffmann et al., 2020). Furin is a type transmembrane protein expressed in tissues and cells, which cleaves the precursors of cell surface receptors and adhesion molecules, but has also a key role as fusion protein of a broad range of viruses such as HIV, Ebola virus, and yellow fever virus (Rockwell et al., 2002; Bestle et al., 2020). Clinical studies showed the effectiveness of camostat mesylate and nafamostat mesylate (Kawase et al., 2012; Hoffmann et al., 2020; Breining et al., 2021). These molecules are cellular serine protease inhibitor (Kawase et al., 2012), which are able to inhibit the TMPRSS2 stopping SARS-CoV-2 infection in lung cells (Hoffmann et al., 2020). Chloroquine has been shown to be

effective against the coronaviruses by different mechanisms of action (Barnard et al., 2006; Gies et al., 2020; Yao et al., 2020). It works as an entry inhibitor by impairing endosomal-mediated entry inhibiting the fusion of the virus to the cell membrane by modulation of the endosomal pH, and prevents acidification, which several viruses use for the fusion and entry process (Vigerust and Shepherd, 2007; Rolain et al., 2007).

### 2.4. N-linked glycosylation inhibitor

Enveloped viruses use cellular glycosylation pathway to modify their biogenesis, antigenicity and infectivity (Vigerust and Shepherd, 2007). The most used type of glycosylation is the N-glycosylation, which begins in the endoplasmic reticulum on a peptide chain. In this reaction, a standard carbohydrate chain is added at the nitrogen atom of a side chain of asparagine (Vigerust and Shepherd, 2007). Studies showed that the N-linked glycosylation on hepatitis C virus (HCV), Ebola (Eichler et al., 2006), Hendra (Bossart et al., 2005), Nipah (Aguilar et al., 2006), and SARS-CoV-1 (Oostra et al., 2006) have key roles in tropism infectivity and immune evasion (Goffard and Dubuisson, 2003). In SARS-CoV-1 and SARS-CoV-2, the surface protein involved in target cell attachment and fusion processes is the spike glycoprotein (S) (Hoffmann et al., 2020; Tripet et al., 2004), a 1255-amino acid precursor polypeptide characterized by 23 potential N-linked glycosylation sites (Xiao et al., 2003). All N-glycosylation sites occur on the amide nitrogen of an asparagine residue (Asn) with exposure of N-acetylglucosamine glycan (GlcNAc) attached to asparagine in  $\beta$  configuration (Stanley et al., 2017). A new therapeutic protocol involves the use of the enzyme asparaginase, which can break down asparagine and inhibits its formation, reducing the synthesis of N-acetylglucosamine molecules (Bellini et al., 2020). Studies showed the antiviral activity of L-asparaginase against HIV-1, retrovirus and herpesvirus infections (Maral and Werner, 1971; Avramis et al., 2001). In this way, the SARS-CoV-2 viral coating and cellular infection are stopped. Chloroquine, or alternatively Hydroxychloroquine, has anti-inflammatory and immunomodulatory activities and can also interfere with the glycosyltransferases's activity (Al-Bari, 2017). It has been shown to be effective *in vitro* and in mouse models against the SARS-CoV-2 by different mechanisms of action (Barnard et al., 2006; Gies et al., 2020; Yao et al., 2020). Chloroquine, as mentioned above, can inhibit the fusion of the virus to the cell membrane by modulation of the endosomal pH (Vigerust and Shepherd, 2007; Rolain et al., 2007) and it can also stop the association between calnexin and calreticulin, misfolding viral proteins (Vigerust and Shepherd, 2007).

## 3. Host-directed therapeutical approaches and against the immune response modulation

Similar to other viruses, coronavirus has the ability to evade the cellular defense (Cheng et al., 2007). Once into the target cell, there may be an inflammatory and apoptotic response followed by a regenerative organ activity or evolves in syndrome (Cheng et al., 2007). The ability to evade the immune system is well represented by the feline coronavirus (FCov). This virus is classified in two serotypes, basing on differences of the S protein amino acid sequence and antibody neutralization (Shiba et al., 2007; Lewis et al., 2015). The two biotypes, referred to feline enteric coronavirus (FECV) and feline infectious peritonitis virus (FIPV), differ for the disease they cause. The FIPV biotype is considered highly virulent and leads to a lethal disease called feline infectious peritonitis (FIP) (Pedersen et al., 1984). The major target cells of FIPV are monocytes and macrophages in which the virus is able to replicate and trigger an activation of these cells (Dewerchin et al., 2005). The infected and activated monocytes express cytokines such as IL-1 $\beta$ , IL-6, IL-10, TNF- $\alpha$  and adhesion molecules (Malbon et al., 2019; Foley et al., 2003) triggering a cytokine storm. Moreover, antibodies enhance the uptake and replication of FIPV in target cells and contribute to a type III hypersensitivity vasculitis (Pedersen, 2009). Although in the cat, an individual

predisposition to the development of FIP has not yet been fully demonstrated, it is conceivable that a genetic risk factors may have a role in disease progression, as has recently been demonstrated in humans ([The Severe Covid-19 GWAS Group, 2020](#)). These recent studies show that genomic regions on chromosome 3 and 9 are associated with severe COVID-19 ([Zeberg and Pääbo, 2020](#)). As happens in the cat with FIP, it is known that the severity of disease caused by coronaviruses is due to the “cytokine storm”. The immune-response in SARS-CoV-1, MERS-CoV, and SARS-CoV-2 is characterized by an excessive production of proinflammatory cytokines such as the interferon gamma-induced protein 10 (IP10), which activates cytotoxic T lymphocytes and monocytes in lung tissue ([Wong et al., 2004](#)). The so called “cytokine storm” can damage the lung, brain, cardiovascular system, gastrointestinal tract, liver, kidney, microcirculation, and eyes ([Bhaskar et al., 2020](#)) resulting in multiorgan dysfunction ([Fig. 2](#)). For this excessive response of the immune system, several drugs with immunomodulatory, anti-inflammatory and immuno-suppressive effect have been proposed. However, their use in diseases caused by coronaviruses in both human and veterinary medicine is still debated. Some molecules have been chosen to activate the shelter while others have observed to block the receptors managed by “harmful” cytokines. Characterized by a repairing and protective role are the following agents: type I interferon alone or in combination by its broad-spectrum antiviral effect ([Cinatl et al., 2003](#)), mesenchymal stem cells which can reduce the inflammatory infiltrate ([Lee et al., 2009](#)), intravenous immunoglobulin ([Jolles et al., 2005](#)), specific neutralizing antibodies ([Zhou and Zhao, 2020](#)) and

human monoclonal antibody such as REGN3048 and eculizumab ([de Wit et al., 2018](#); [Diurno et al., 2020](#)). Agents proposed to manage “harmful” cytokines are: anti-interleukin-6 receptor such as sarilumab ([Rose-John, 2012](#)), interleukin-1 (IL-1) receptor antagonists, such as anakinra ([McCreary and Pogue, 2020](#)), and thalidomide which decreases the synthesis of TNF-alpha ([Zhu et al., 2014](#)). Corticosteroids are known to reduce the numbers of CD4 and CD8 T cells and cytokine levels inducing immunosuppression. Studies suggested the role of corticosteroid treatment in SARS patients. These drugs can reduce proinflammatory cytokines playing a major role in lung immunopathology, however corticosteroids can induce immunosuppression leading to enhanced respiratory disease or increased CoV shedding ([McCreary and Pogue, 2020](#); [Veronese et al., 2020](#)). Even in cats with FIP, both anti-inflammatory and immuno-suppressive drugs such as prednisolone, alkylating drugs (cy-clophosphamide), and inhibitors of specific cytokines (pentoxifylline) have been used to reduce clinical signs, but without evidence about the disease outcome ([Pedersen, 2014](#)). In a study conducted in 2017, the Polyprenyl Immunostimulant was proposed as treatment for the reduction of clinical signs in sixty cats affected by FIP ([Legendre et al., 2017](#)). This drug can upregulate Th-1 type pathway via toll-like receptors, however, there have been no further studies demonstrating its benefit. On the basis of some preliminary case reports, showing a mild disease course in COVID-19 affected patients, maintained in a regimen of immunosuppression by calcineurin inhibitors (CNIs) ([Zhu et al., 2020](#); [Guillen et al., 2020](#); [D’Antiga, 2020](#); [Monti et al., 2020](#)), such as cyclosporine (Cys) and

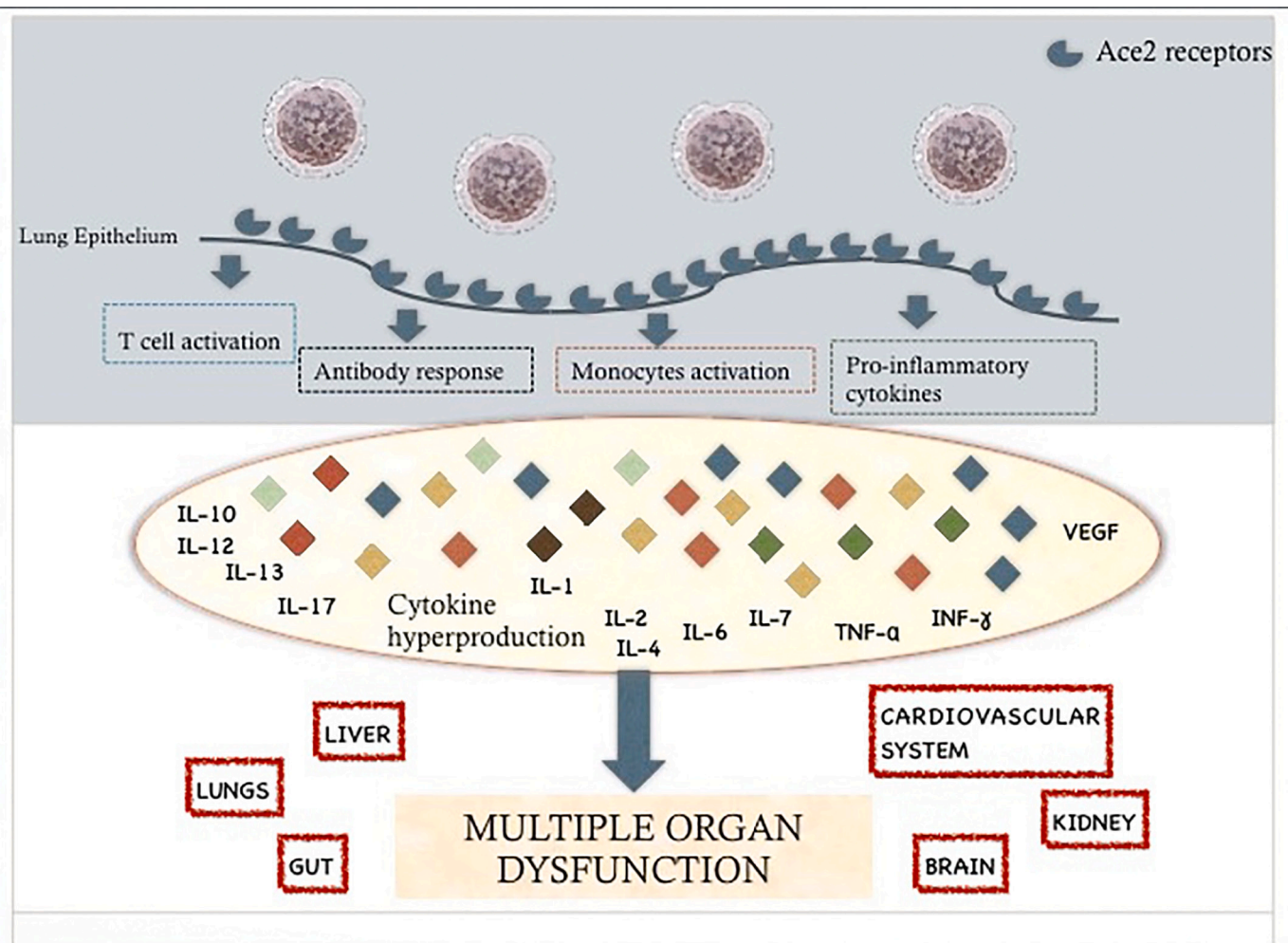


Fig. 2. Schematic representation of the “cytokines storm”.

tacrolimus (TAC), which are currently used in the setting of transplantation (Kuypers, 2020) and Rheumatic Disorders (RMDs) (Cavagna et al., 2020b), different studies were performed to evaluate also a potential antiviral activity of these molecules (Cavagna et al., 2020a). On the other hand, immunosuppressive treatments are being studied as possible therapeutic options in the hyper-inflammatory phase of the COVID-19 (Mehta et al., 2020). The antiviral effect of CNIs has been evidenced in vitro against some coronavirus strains (Pfefferle et al., 2011; Ma-Lauer et al., 2020; Carbajo-Lozoya et al., 2012; de Wilde et al., 2011). Clinical evidences of COVID-19 course in patients treated with CNIs alone or in association with other immune-suppressants (ISs) appears to be generally mild, without development of superimposed infections despite immunosuppression, and that also in the case of SARS-COV-2 related lung involvement or of previously diagnosed interstitial lung disease (ILD). Substantially, CNIs based chronic immunosuppressive regimens do not increase the risk of severe course of COVID-19 or complications, decreasing the mortality rate (Cavagna et al., 2020a). The CNIs based chronic immunosuppressive regimens influences the clinical course of the disease preventing the occurrence of the huge alveolar macrophage activation, with consequent release of pro-inflammatory cytokines that has been described in the context of SARS-COV-2 (Mehta et al., 2020). In fact, although high sensitivity C-reactive protein (hs-CRP) levels were similar between confirmed COVID-19 with and without immunosuppression, IL-6 levels were much lower in the immunosuppressed group. Interestingly, both Cys and TAC inhibit viral replication in a number of CoV strains, including SARS-CoV-1, through the inhibition of peptidyl-prolyl cis-trans isomerases, such as cyclophilin A and FK506-binding proteins, that are cellular interaction partners of CoV non-structural protein 1 (Nsp1) (Pfefferle et al., 2011; Ma-Lauer et al., 2020; Carbajo-Lozoya et al., 2012). In this sense, it is conceivable that Cys and TAC exert antiviral activity also towards SARS-CoV-2. On this basis, it is reasonable to add also CNIs in the list of ISS possible therapeutic options for COVID-19 (Sarzi-Puttini et al., 2020).

#### 4. New approaches of bacteriotherapy

The live microbes, such as probiotics, possess health benefits on the host when administered in appropriately adequate amount. Several studies showed their role in, stimulating mucosal barrier function and modulating the immune system (Brown and Valiere, 2004) exerting beneficial effects through modulation of vitamin D (Li et al., 2015). Similarly to other *Coronaviridae*, as FECoV in the cat, ferret enteric CoV (FRECV), and CRCoV in dogs, the invasive process of SARS-COV-2 is due to enzymes which are linked to intestinal epithelial cells (IECs). Coronaviruses constantly change their binding patterns; the potential target in the lungs (as in primates or in *Mustelidae* or *Felidae*) or macrophages (mainly in *Felidae*) can change, but not in the small intestine, where it remains constant. The IECs could be a reservoir for coronaviruses (Feng et al., 2020). Different studies show that some strains of *Lactobacilli* and *Bifidobacteria* could inhibit viruses such as influenza virus, rhinovirus, respiratory syncytial virus, adenovirus, and pneumovirus (Leyer et al., 2009; Li et al., 2019). Chinese researchers have investigated changes in the microbiota in patients died by Covid-19, showing a significant decrease in *Bifidobacteria* and *Lactobacilli*, and an increase in opportunistic bacteria such as *Corynebacterium* or *Ruthenibacterium*. The severity of hypoxemia was strongly correlated with high levels of immune cells and markers of inflammation. In addition, during the acute respiratory distress syndrome (ARDS) of affected patients, a decrease in the tissue oxygenation, also in GI tract, can alter the composition of the gut microbiota decreasing proportion of oxygen-tolerant organisms and increasing anaerobic phyla, as *Clostridiales* (Albenberg et al., 2014).

The link between hyper inflammation and intestinal dysbiosis appears to be a high risk for ARDS. In the acute phase, virus cDNA was detected in blood in 10% of patients and in stool of 50% of patients, suggesting that feces could be a mode of contamination (Leung et al., 2003). The gut involvement might explain the wide variation in viral

load from one test to another in the same patients (Yu et al., 2020). Recently, studies demonstrated that the use of oral bacteriotherapy could be an option to treat COVID-19. A select mixture of bacteria, previously demonstrated with antiviral activity (Li et al., 2019), was administered with anti-COVID-19 treatment (chloroquine, antibiotics, and/or tocilizumab) and compared to other COVID-19 positive treated subjects, without bacteriotherapy (Ceccarelli et al., 2020; D'Ettoire et al., 2020). Results suggested a different surviving rate (four deaths vs. zero death in probiotic treated patients), and a lower estimated risk to develop respiratory failure during COVID-19 course (D'Ettoire et al., 2020). Also for the other signs and symptoms associated with COVID-19, i.e. diarrhea, fever, cough, dyspnea, asthenia, and myalgia a significant improvement is already evident as early as after 24 hours after the start of the bacteriotherapy, and seventy-two hours after the start of the bacteriotherapy, the 100% of treated patients showed a remission of symptoms vs. about the 45% of the control group (D'Ettoire et al., 2020). These results suggest a possibility important role of the “gut-lung axis” in the control of the COVID-19 infection (Enaud et al., 2020; Dumas et al., 2018). Studies show that during corona- and influenza-virus infections, certain bacteria (generally opportunistic or pathogen species) can support the viral activation either by secreting proteases that cleave Spikes or influenza hemagglutinin (HA) or due to activation of cellular proteases, possibly contributing to the intracellular virus entry (Böttcheri-Friebertshäuser et al., 2013). In this sense, an oral bacteriotherapy could reduce this viral “priming” at intestinal level. There are potential anatomical communications and complex pathways involving intestine and lungs (GLA) (Enaud et al., 2020; Dumas et al., 2018). The mesenteric lymphatic system is the pathway between the lungs and the intestine, through which bacteria or fragments or metabolites, can cross the intestinal barrier to reach systemic circulation and influence the pulmonary immune response (Bingula et al., 2017; McAleer and Kolls, 2018; Trompette et al., 2014). It has been showed that mice with SCFAs receptors deficiency show increased inflammatory responses in experimental models of asthma (Trompette et al., 2014). Short chain fatty acids (SCFAs), produced primarily by bacterial fermentation of dietary fiber, act in the lungs as signaling to attenuate inflammatory and allergic responses (Cait et al., 2018; Anand and Mande, 2018). Additionally, oral bacteriotherapy enhance the cellular antioxidative defense systems protecting against reactive oxygen species (ROS) generated by viruses (Hosakote et al., 2011). By using selected bacterial strains, enhancing the production of both the nuclear factor erythroid 2p45-related factor 2 (Nrf2) and its target Heme oxygenase-1 (HO-1) (Castelli et al., 2020), an antiviral effect was induced through a reduction of oxidative stress. Nrf2 and HO-1 have significant antiviral activity against viruses, including HIV, hepatitis B virus, influenza virus, respiratory syncytial virus, dengue virus, and Ebola virus among others (Devadas and Dhawan, 2006; Protzer et al., 2007; Hashiba et al., 2001; Ma et al., 2016; Janyra et al., 2017; Tseng et al., 2016; Hill-Batorski et al., 2013). Notably, beneficial properties of HO-1 expression have been reported for viruses responsible for lung disease, as demonstrated in a Mice model in which an overexpressed HO-1 in the lungs lead to a lower inflammatory cells infiltration, decreased apoptosis of respiratory epithelial cells, preventing an exacerbated immune response in this tissue, and subsequent damage (Hashiba et al., 2001). As mentioned above, the cytokine storm is an offensive inflammatory response resulting from COVID-19 infection characterized with a hyper-production of pro-inflammatory molecules. Probiotics can exert functional roles in preserving the equilibrium between innate and adaptive immune response. However, there is still little scientific evidence on the usefulness of bacteriotherapy in coronavirus infections in human and veterinary medicine.

#### 5. Conclusion

Coronaviruses are showing a high ability to mutate and jump the animal-human barrier, acquiring over time the zoonotic role. As shown

by coronaviruses in animals, this pathogen can often cause a fatal disease. In the cat, the virulent biotype of FCoV results in lethal inflammation, associated with systemic and neurological disorders; in the dog, the virus can infect the upper respiratory gastro-enteric tracts leading to fatal disease; in the pig, the porcine CoV causes significant morbidity and mortality of piglets due to enteric and nervous system infection; in the cattle, the infection by Bovine CoVs causes severe or fatal infection (Weiss and Navas-Martin, 2005; Amer, 2018). Within the “One Health concept”, it is necessary to improve the knowledge and the study of their pathogenic role in both human and animals. Different therapeutic protocols have been proposed during the global outbreaks of SARS-CoV-1, MERS-CoV and SARS-CoV-2. These approaches can counteract the viral entry phase, as a target, or can act modulating the cytokines storm aggravating clinical symptoms. However, a definitive protocol has not yet been reached. However, to date, the therapeutic protocols proposed for both animal and human coronaviruses are a palliative to limit the worsening of the disease. Furthermore, even if experimental studies have shown the efficacy of antivirals against feline coronavirus (Pedersen et al., 2019; Dickinson et al., 2020), it cannot be excluded that coronaviruses are able to rapidly acquire resistance factors to specific antiviral drugs. In author’s opinion the use of probiotics and postbiotics products can be an interesting option in the management of patients hospitalized for coronavirus spp. infection, both human and animals.

## Declaration of Competing Interest

The authors declare no conflict of interest.

## References

- Addie, D.D., Curran, S., Bellini, F., Crowe, B., Sheehan, E., Ukrainchuk, L., Decaro, N., 2020. Oral Mutian®X stopped faecal feline coronavirus shedding by naturally infected cats. *Res. Vet. Sci.* 130, 222–229.
- Agostini, M.L., Andres, E.L., Sims, A.C., Graham, R.L., Sheahan, T.P., Lu, X., Smith, E.C., Case, J.B., Feng, J.Y., Jordan, R., Ray, A.S., Cihlar, T., Siegel, D., Mackman, R.L., Clarke, M.O., Baric, R.S., Denison, M.R., 2018. Coronavirus susceptibility to the antiviral remdesivir (GS-5734) is mediated by the viral polymerase and the proofreading exoribonuclease. *MBio* 9 (2) (e00221-18).
- Aguiar, H.C., Matreyek, K.A., Filone, C.M., Hashimi, S.T., Levroney, E.L., Negrete, O.A., Bertolotti-Ciarlet, A., Choi, D.Y., McHardy, I., Fulcher, J.A., Su, S.V., Wolf, M.C., Kohatsu, L., Baum, L.G., Lee, B., 2006. N-glycans on Nipah virus fusion protein protect against neutralization but reduce membrane fusion and viral entry. *J. Virol.* 80, 4878–4889.
- Al-Bari, M., 2017. Targeting endosomal acidification by chloroquine analogs as a promising strategy for the treatment of emerging viral diseases. *Pharmacol. Res. Perspect.* 5 (1), e00293.
- Albenberg, L., Esipova, T.V., Judge, C.P., Bittinger, K., Chen, J., Laughlin, A., Grunberg, S., Baldassano, R.N., Lewis, J.D., Li, H., Thom, S.R., Bushman, F.D., Vinogradov, S.A., Wu, G.D., 2014. Correlation between intraluminal oxygen gradient and radial partitioning of intestinal microbiota in humans and mice. *Gastroenterology* 147 (5) (1055–1063.e8).
- Amer, H.M., 2018. Bovine-like coronaviruses in domestic and wild ruminants. *Anim. Health Res. Rev.* 19, 113–124.
- Anand, S., Mande, S.S., 2018. Diet, microbiota and gut-lung connection. *Front. Microbiol.* 9, 2147.
- Avramis, V.I., Kwock, R., Avramis, I.A., Cohen, L.J., Inderlied, C., 2001. Synergistic antiviral effect of PEG-Asparaginase (ONCASPARG), with protease inhibitor alone and in combination with RT inhibitors against HIV-1 infected T-cells: A model of HIV-1-induced T-cell lymphoma. *In vivo* (Athens, Greece) 15 (1), 1–9V.
- Barnard, D.L., Hubbard, V.D., Burton, J., Smees, D.F., Morrey, J.D., Otto, M.J., Sidwell, R.W., 2004. Inhibition of severe acute respiratory syndrome-associated coronavirus (SARSCoV) by calpain inhibitors and beta-d-N4-hydroxycytidine. *Antivir. Chem. Chemother.* 15, 15–22.
- Barnard, D.L., Day, C.W., Bailey, K., Heiner, M., Montgomery, R., Lauridsen, L., Chan, P.K., Sidwell, R.W., 2006. Evaluation of immunomodulators, interferons and known in vitro SARS-coV inhibitors for inhibition of SARS-coV replication in BALB/c mice. *Antivir. Chem. Chemother.* 17, 275–284.
- Bellini, F., Rossi, G., Piergallini, R., inventors; Composition and methods for treatment of Coronaviridae infections. U.S. Patent and Trademark; Application n° 63013633, EFS ID: 39228068, Confirmation number 7730, Attorney Docket n° 105029-003, Rept date 22.04.2020 – 2020 <https://www.uspto.gov/patent>.
- Bestle, D., Heindl, M.R., Limburg, H., Van Lam van, T., Pilgram, O., Moulton, H., Stein, D.A., Harges, K., Eickmann, M., Dolnik, O., Rohde, C., Klenk, H. D., Garten, W., Steinmetzer, T., & Böttcher-Friebertshäuser, E., 2020. TMPRSS2 and furin are both essential for proteolytic activation of SARS-CoV-2 in human airway cells. *Life Sci.* 3 (9), e202000786.
- Bhaskar, S., Sinha, A., Banach, M., Mittoo, S., Weissert, R., Kass, J.S., Rajagopal, S., Pai, A.R., Kutty, S., 2020. Cytokine storm in COVID-19 Immunopathological mechanisms, clinical considerations, and therapeutic approaches: The REPROGRAM Consortium Position Paper. *Front. Immunol.* 11, 1648.
- Bingula, R., Filaire, M., Radosevic-Robin, N., Bey, M., Berthon, J.-Y., Bernalier-Donadille, A., Vasson, M.P., Filaire, E., 2017. Desired turbulence? Gut-lung axis, immunity, and lung cancer. *J. Oncol.* 5035371.
- Bossart, K.N., Crameri, G., Dimitrov, A.S., Mungall, B.A., Feng, Y.R., Patch, J.R., Choudhary, A., Wang, L.F., Eaton, B.T., Broder, C.C., 2005. Receptor binding, fusion inhibition, and induction of cross-reactive neutralizing antibodies by a soluble G glycoprotein of Hendra virus. *J. Virol.* 79, 6690–6702.
- Böttcher, E., Matrosovich, T., Beyerle, M., Klenk, H.D., Garten, W., Matrosovich, M., 2006. Proteolytic activation of influenza viruses by serine proteases TMPRSS2 and HAT from human airway epithelium. *J. Virol.* 80 (19), 9896–9898.
- Böttcher-Friebertshäuser, E., Klenk, H.D., Garten, W., 2013. Activation of influenza viruses by proteases from host cells and bacteria in the human airway epithelium. *Pathog. Dis.* 69, 87–100.
- Breining, P., Frølund, A.L., Højen, J.F., Gunst, J.D., Staerke, N.B., Saedder, E., Cases-Thomas, M., Little, P., Nielsen, L.P., Sogaard, O.S., Kjolby, M., 2021. Camostat mesylate against SARS-CoV-2 and COVID-19 - Rationale, dosing and safety. *Basic Clin. Pharmacol. Toxicol.* 128 (2), 204–212.
- Brown, A.C., Valiere, A., 2004. Probiotics and medical nutrition therapy. *Nutr. Clin. Care.* 7, 56–68.
- Cait, A., Hughes, M.R., Antignano, F., Cait, J., Dimitriu, P.A., Maas, K.R., Reynolds, L.A., Hacker, L., Mohr, J., Finlay, B.B., Zaph, C., McNagny, K.M., Mohn, W.W., 2018. Microbiome-driven allergic lung inflammation is ameliorated by short-chain fatty acids. *Mucosal Immunol.* 11, 785–795.
- Carbajo-Lozoya, J., Müller, M.A., Kallies, S., Thiel, V., Drosten, C., von Brunn, A., 2012. Replication of human coronaviruses SARS-CoV, HCoVNL63 and HCoV-229E is inhibited by the drug FK506. *Virus Res.* 165, 112–117.
- Castelli, V., d’Angelo, M., Lombardi, F., Alfonso, M., Antonosante, A., Catanesi, M., Benedetti, E., Palumbo, P., Cifone, M.G., Giordano, A., Desideri, G., Cimini, A., 2020. Effects of the probiotic formulation SLAB51 in in vitro and in vivo Parkinson’s disease models. *Aging* 12, 4641–4659.
- Cavagna, L., Bruno, R., Zanframundo, G., Gregorini, M., Seminari, E., Di Matteo, A., Rampino, T., Montecucco, C., Pelenghi, S., Cattadori, B., Pattonieri, E.F., Vitello, P., Bertani, A., Sambataro, G., Vancheri, C., Bonetto, V., Monti, M.C., Ticozzelli, E., Turco, A., Oggionni, T., Corsico, A., Codullo, V., Morosini, M., Gneccchi, M., Pellegrini, C., Meloni, F., 2020a. Clinical presentation and evolution of COVID-19 in immunosuppressed patients. Preliminary evaluation in a North Italian cohort on calcineurin-inhibitors based therapy. *medRxiv*. <https://doi.org/10.1101/2020.04.26.20080663> preprint. (version posted May 1, 2020).
- Cavagna, L., Caporali, R., Abdi-Ali, L., Dore, R., Meloni, F., Montecucco, C., 2020b. Cyclosporine in anti-Jo1-positive patients with corticosteroid-refractory interstitial lung disease. *J. Rheumatol.* 40, 484–492.
- CDC, 2019. Centers for Disease Control and Prevention. About MERS. Centers for Disease Control and Prevention.
- Ceccarelli, G., Scagnolari, C., Pugliese, F., Mastroianni, C.M., d’Ettorre, G., 2020. COVID-19 and probiotics. *Lancet Gastroenterol.* 5, 721–722.
- Chan, J.F., Chan, K.H., Kao, R.Y., To, K.K., Zheng, B.J., Li, C.P., Li, P.T., Dai, J., Mok, F.K., Chen, H., Hayden, F.G., Yuen, K.Y., 2013a. Broad-spectrum antivirals for the emerging Middle East respiratory syndrome coronavirus. *J. Infect.* 67, 606–616.
- Chan, J.F., To, K.K., Tse, H., Jin, D.Y., Yuen, K.Y., 2013b. Interspecies transmission and emergence of novel viruses: Lessons from bats and birds. *Trends Microbiol.* 21 (10), 544–555.
- Chandwani, A., Shuter, J., 2008. Lopinavir/ritonavir in the treatment of HIV-1 infection: a review. *Ther. Clin. Risk Manag.* 4 (5), 1023–1033.
- Cheng, V.C.C., Lau, S.K.P., Woo, P.C.Y., Yuen, K.Y., 2007. Severe acute respiratory syndrome coronavirus as an agent of emerging and reemerging infection. *Clinical Microbiol. Rev.* 20, 660–694.
- Cinatli, J., Morgenstern, B., Bauer, G., Chandra, P., Rabenau, H., Doerr, H.W., 2003. Treatment of SARS with human interferons. *Lancet.* 362 (9380), 293–294.
- Cvetkovic, R.S., Goa, K.L., 2003. Lopinavir/Ritonavir. *Drugs.* 63, 769–802.
- D’Ettorre, G., Giancarlo, C., Marazzato, M., Campagna, G., Pinacchio, C., Alessandri, F., Ruberto, F., Rossi, G., Celani, L., Scagnolari, C., Mastropietro, C., Trinchieri, V., Recchia, G.E., Mauro, V., Antonelli, G., Pugliese, F., Mastroianni, C.M., 2020. Challenges in the management of SARS-CoV2 infection: the role of oral bacteriotherapy as complementary therapeutic strategy to avoid the progression of COVID-19. *Front. Med.* 7, 389.
- D’Antiga, L., 2020. Coronaviruses and immunosuppressed patients. The facts during the third epidemic. *Liver Transplant.* 26 (6), 832–834.
- Dayer, M.R., Taleb-Gassabi, S., Dayer, M.S., 2017. Lopinavir; a potent drug against coronavirus infection: Insight from molecular docking study. *Arch. Clin. Infect. Dis.* 12, e13823.
- De Wilde, A.H., Zevenhoven-Dobbe, J.C., van der Meer, Y., Thiel, V., Narayanan, K., Makino, S., Snijder, E.J., van Hemertcorresponding, M.J., 2011. Cyclosporin A inhibits the replication of diverse coronaviruses. *J. Gen. Virol.* 92, 2542–2548.
- de Wit, E., Feldmann, F., Okumura, A., Horne, E., Haddock, E., Saturday, G., Scott, D., Erlandson, K.J., Stahl, N., Lipsich, L., Kyrtasous, C.A., Feldmann, H., 2018. Prophylactic and therapeutic efficacy of mAb treatment against MERS-CoV in common marmosets. *Antiviral Res.* 156, 64–71.
- Decaro, N., Lorusso, A., 2020. Novel human coronavirus (SARS-CoV-2): A lesson from animal coronaviruses. *Vet. Microbiol.* 244, 108693.
- Devadas, K., Dhawan, S., 2006. Hemin activation ameliorates HIV-1 infection via heme oxygenase-1 induction. *J. Immunol.* 176, 4252–4257.

- Dewerchin, H.L., Cornelissen, E., Nauwynck, H.J., 2005. Replication of feline coronaviruses in peripheral blood monocytes. *Arch. Virol.* 150, 2483–2500.
- Di Teodoro, G., Valleriani, F., Puglia, I., Monaco, F., Di Pancrazio, C., Luciani, M., Krasteva, I., Petrini, A., Maracci, M., D'Alterio, N., Curini, V., Iorio, M., Migliorati, G., Di Domenico, M., Morelli, D., Calistri, P., Savini, G., Decaro, N., Holmes, E.C., Lorusso, A., 2020. SARS-CoV-2 replicates in respiratory ex vivo organ cultures of domestic ruminant species. *Vet. Microbiol.* 24 (252), 108933.
- Dickinson, P.J., Bannasch, M., Thomas, S.M., Murthy, V.D., Vernau, K.M., Liepnieks, M., Montgomery, E., Knickelbein, K.E., Murphy, B., Pedersen, N.C., 2020. Antiviral treatment using the adenosine nucleoside analogue GS-441524 in cats with clinically diagnosed neurological feline infectious peritonitis. *J. Vet. Intern. Med.* 34 (4), 1587–1593.
- Diurno, F., Numis, F.G., Porta, G., Cirillo, F., Maddaluno, S., Ragozzino, A., De Negri, P., Di Gennaro, C., Pagano, A., Allegorico, E., Bressy, L., Bosso, G., Ferrara, A., Serra, C., Montisci, A., D'Amico, M., Schiano Lo Morello, S., Di Costanzo, G., Tucci, A.G., Marchetti, P., Di Vincenzo, U., Sorrentino, I., Casciotta, A., Fusco, M., Buonerba, C., Berretta, M., Ceccarelli, M., Nunnari, G., Diessa, Y., Cicala, S., Facchini, G., 2020. Eculizumab treatment in patients with COVID-19: preliminary results from real life ASL Napoli 2 Nord experience. *Eur. Rev. Med. Pharmacol. Sci.* 24 (7), 4040–4047.
- Dumas, A., Bernard, L., Poquet, Y., Lugo-Villarino, G., Neyrolles, O., 2018. The role of the lung microbiota and the gut-lung axis in respiratory infectious diseases. *Cell. Microbiol.* 20, e12966.
- Dyall, J., Coleman, C.M., Hart, B.G., Venkataraman, T., Holbrook, M.R., Kindrachuk, J., Johnson, R.F., Olinger Jr., G.G., Jahrling, B.B., Laidlaw, M., Johansen, L.M., Lear-Rooney, C.M., Glass, P.J., Hensley, L.E., Frieman, M.B., 2014. Repurposing of clinically developed drugs for treatment of middle east respiratory syndrome coronavirus infection. *Antimicrob. Agents Chemother.* 58, 4885–4893.
- Eichler, R., Lenz, O., Garten, W., Strecker, T., 2006. The role of single N-glycans in proteolytic processing and cell surface transport of the Lassa virus glycoprotein GP-C. *Virol. J.* 3, 41.
- Elfiky, A.A., Mahdy, S.M., Elshemy, W.M., 2017. Quantitative structure-activity relationship and molecular docking revealed a potency of anti-hepatitis C virus drugs against human corona viruses. *J. Med. Virol.* 89, 1040–1047.
- Enaud, R., Prevel, R., Ciarlo, E., Beaufils, F., Wiecek, G., Guery, B., Delhaes, L., 2020. The gut-lung axis in health and respiratory diseases: a place for inter-organ and inter-kingdom crosstalks. *Front. Cell. Infect. Microbiol.* 10, 9.
- Feng, Z., Wang, Y., Qi, W., 2020. The Small Intestine, an Underestimated Site of SARS-CoV-2 Infection: From Red Queen Effect to Probiotics (Preprints 2020, 2020030161).
- Foley, J.E., Rand, C., Leutenegger, C., 2003. Inflammation and changes in cytokine levels in neurological feline infectious peritonitis. *J. Feline Med. Surg.* 5 (6), 313–322.
- Gao, Y., Yan, L., Huang, Y., Liu, F., Zhao, Y., Cao, L., Wang, T., Sun, Q., Ming, Z., Zhang, L., Ge, J., Zheng, L., Zhang, Y., Wang, H., Zhu, Y., Zhu, C., Hu, T., Hua, T., Zhang, B., Yang, X., Li, J., Yang, H., Liu, Z., Xu, W., Guddat, L.W., Wang, Q., Lou, Z., Rao, Z., 2020. Structure of the RNA-dependent RNA polymerase from COVID-19 virus. *Science*. <https://doi.org/10.1126/science.abb7498>.
- Gies, V., Bekaddour, N., Dieudonné, Y., Guffroy, A., Frenger, Q., Gros, F., Rodero, M.P., Herbeuval, J.P., Korganow, A.S., 2020. Beyond anti-viral effects of chloroquine/hydroxychloroquine. *Front. Immunol.* 2 (11), 1409.
- Glowska, I., Bertram, S., Müller, M.A., Allen, P., Soilleux, E., Pfefferle, S., Steffen, I., Tsegaye, T.S., He, Y., Gnriss, K., Niemeier, D., Schneider, H., Drosten, C., Pöhlmann, S., 2011. Evidence that TMPRSS2 activates the severe acute respiratory syndrome coronavirus spike protein for membrane fusion and reduces viral control by the humoral immune response. *J. Virol.* 85 (9), 4122–4134.
- Goffard, A., Dubuisson, J., 2003. Glycosylation of hepatitis C virus envelope proteins. *Biochimie* 85, 295–301.
- Guillen, E., Pineiro, G.J., Revuelta, I., Rodriguez, D., Bodro, M., Moreno, A., Campistol, J.M., Diekmann, F., Ventura-Aguar, P., 2020. Case report of COVID-19 in a kidney transplant recipient: Does immunosuppression alter the clinical presentation? *Am. J. Transplant.* 20 (7), 1875–1878.
- Hamming, I., Timens, W., Bulthuis, M.L., Lely, A.T., Navis, G., van Goor, H., 2004. Tissue distribution of ACE2 protein, the functional receptor for SARS coronavirus. A first step in understanding SARS pathogenesis. *J. Pathol.* 203 (2), 631–637.
- Hashiba, T., Suzuki, M., Nagashima, Y., Suzuki, S., Inoue, S., Tsuburai, T., Matsuse, T., Ishigatubo, Y., 2001. Adenovirus-mediated transfer of heme oxygenase-1 cDNA attenuates severe lung injury induced by the influenza virus in mice. *Gene Ther.* 8, 1499–1507.
- Heald-Sargent, T., Gallagher, T., 2012. Ready, set, fuse! The coronavirus spike protein and acquisition of fusion competence. *Viruses* 4 (4), 557–580.
- Hill-Batorski, L., Halfmann, P., Neumann, G., Kawaoka, Y., 2013. The cytoprotective enzyme heme oxygenase-1 suppresses Ebola virus replication. *J. Virol.* 87 (24), 13795–13802.
- Hoffmann, M., Kleine-Weber, H., Schroeder, S., Krüger, N., Herrler, T., Erichsen, S., Schiergens, T.S., Herrler, G., Wu, N.H., Nitsche, A., Müller, M.A., Drosten, C., Pöhlmann, S., 2020. SARS-CoV-2 cell entry depends on ACE2 and TMPRSS2 and is blocked by a clinically proven protease inhibitor. *Cell* 181 (2) (271–280.e8).
- Hosakote, Y.M., Jantzi, P.D., Esham, D.L., Spratt, H., Kurosky, A., Casola, A., Garofalo, R. P., 2011. Viral-mediated inhibition of antioxidant enzymes contributes to the pathogenesis of severe respiratory syncytial virus bronchiolitis. *Am. J. Respir. Crit. Care Med.* 183 (11), 1550–1560.
- Hung, I.F.N., Lung, K.C., Tso, E.Y., Liu, R., Chung, T.W., Chu, M.Y., Ng, Y.Y., Lo, J., Chan, J., et al., 2020. Triple combination of interferon beta-1b, lopinavir-ritonavir, and ribavirin in the treatment of patients admitted to hospital with COVID-19: an open-label, randomised, phase 2 trial. *Lancet* 395 (10238), 1695–1704.
- Janyra, A., Espinoza, M.A., León, P.F., Céspedes, R.S., Gómez Canedo-Marroquín, G., Riquelme, S.A., Salazar-Echeagarai, F.J., Blancou, P., Simon, T., Anegón, I., Lay, M.K., González, P.A., Riedel, C.A., Bueno, S.M., Kalergis, A.M., 2017. Heme oxygenase-1 modulates human respiratory syncytial virus replication and lung pathogenesis during infection. *J. Immunol.* 199 (1), 212–223.
- Jin, Y.H., Cai, L., Cheng, H., Cheng, Z.S., Cheng, H., Deng, T., Fan, Y.P., Fang, C., Huang, D., Huang, L.Q., Huang, Q., Han, Y., Hu, B., Hu, F., Li, B.H., Li, Y.R., Liang, K., Lin, L.K., Luo, L.S., Ma, J., Ma, L.L., Peng, Z.Y., Pan, Y.B., Pan, Z.Y., Ren, X.Q., Sun, H.M., Wang, Y., Wang, Y.Y., Weng, H., Wei, C.J., Wu, D.F., Xia, J., Xiong, Y., Xu, H.B., Yao, X.M., Yuan, Y.F., Ye, T.S., Zhang, X.C., Zhang, Y.W., Zhang, Y.G., Zhang, H.M., Zhao, Y., Zhao, M.J., Zi, H., Zeng, X.T., Wang, Y.Y., Wang, X.H., 2020. A rapid advice guideline for the diagnosis and treatment of 2019 novel coronavirus (2019-nCoV) infected pneumonia (standard version). *Mil. Med. Res.* 7 (1), 4.
- Jolles, S., Sewell, W.A., Misbah, S.A., 2005. Clinical uses of intravenous immunoglobulin. *Clin. Exp. Immunol.* 142 (1), 1–11.
- Kawase, M., Shirato, K., van der Hoek, L., Taguchi, F., Matsuyama, S., 2012. Simultaneous treatment of human bronchial epithelial cells with serine and cysteine protease inhibitors prevents severe acute respiratory syndrome coronavirus entry. *J. Virol.* 86, 6537–6545.
- Kiliani, A., Baker, S.C., 2014. Cell-based antiviral screening against coronaviruses: developing virus-specific and broad-spectrum inhibitors. *Antiviral Res.* 101, 105–112.
- Kim, Y., Shivanna, V., Narayanan, S., 2015. Broad-spectrum inhibitors against 3C-like proteases of feline coronaviruses and feline caliciviruses. *J. Virol.* 89, 4942–4950.
- Kuypers, D.R.J., 2020. Inpatient variability of tacrolimus exposure in solid organ transplantation: a novel marker for clinical outcome. *Clin. Pharmacol. Ther.* 107, 347–358.
- Lee, J.W., Gupta, N., Serikov, V., Matthay, M.A., 2009. Potential application of mesenchymal stem cells in acute lung injury. *Expert Opin. Biol.* 9, 1259–1270.
- Legendre, A.M., Kuritz, T., Galyon, G., Baylor, V.M., Heidel, R.E., 2017. Polypropyl immunostimulant treatment of cats with presumptive non-effusive feline infectious peritonitis in a field study. *Front. Vet. Sci.* 4, 7.
- Leung, W.K., To, K.F., Chan, P.K., Chan, H.L., Wu, A.K., Lee, N., Yuen, K.Y., Sung, J.J., 2003. Enteric involvement of severe acute respiratory syndrome-associated coronavirus infection. *Gastroenterol.* 125 (4), 1011–1017.
- Lewis, C.S., Porter, E., Matthews, D., Kipar, A., Tasker, S., Helps, C.R., Siddell, S.G., 2015. Genotyping coronaviruses associated with feline infectious peritonitis. *J. Gen. Virol.* 96, 1358–1368.
- Leyer, G.J., Li, S., Mubasher, M.E., Reifer, C., Ouwehand, A.C., 2009. Ouwehand probiotic effects on cold and influenza-like symptom incidence and duration in children. *Pediatrics* 124 (2), e172–e179.
- Li, F., 2016. Structure, function, and evolution of coronavirus spike proteins. *Annu. Rev. Virol.* 3 (1), 237–261.
- Li, W., Moore, M.J., Vasilieva, N., Sui, J., Wong, S.K., Berne, M.A., Somasundaran, M., Sullivan, J.L., Luzuriaga, K., Greenough, T.C., Choe, H., Farzan, M., 2003. Angiotensin-converting enzyme 2 is a functional receptor for the SARS coronavirus. *Nature*. 426, 450–454.
- Li, Y.C., Chen, Y., Du, J., 2015. Critical roles of intestinal epithelial vitamin D receptor signaling in controlling gut mucosal inflammation. *J. Steroid. Biochem. Mol. Biol.* 148, 179–183.
- Li, N., Ma, W.T., Pang, M., Fan, Q.L., Hua, J.L., 2019. The commensal microbiota and viral infection: a comprehensive review. *Front. Immunol.* 10, 1551.
- Lo, M.K., Jordan, R., Arvey, A., Sudhamsu, J., Shrivastava-Ranjan, P., Hotard, A.L., Flint, M., McMullan, L.K., Siegel, D., Clarke, M.O., Mackman, R.L., Hui, H.C., Perron, M., Ray, A.S., Cihlar, T., Nichol, S.T., Spiropoulou, C.F., 2017. GS-5734 and its parent nucleoside analog inhibit Filo-, Pneumo-, and Paramyxoviruses. *Sci. Rep.* 7, 43395.
- Luan, B., Huynh, T., Cheng, X., Lan, G., Wang, H.R., 2020. Targeting proteases for treating COVID-19. *J. Proteome Res.* 19 (11), 4316–4326.
- Ma, L.L., Wang, H.Q., Wu, P., Hu, J., Yin, J.Q., Wu, S., Ge, M., Sun, W.F., Zhao, J.Y., Aisa, H.A., Li, Y.H., Jiang, J.D., 2016. Rupestonic acid derivative YZH-106 suppresses influenza virus replication by activation of heme oxygenase-1-mediated interferon response. *Free Radic. Biol. Med.* 96, 347–361.
- Ma-Lauer, Y., Zheng, Y., Malešević, M., von Brunn, B., Fischer, G., von Brunn, A., 2020. Influences of cyclosporin A and non-immunosuppressive derivatives on cellular cyclophilins and viral nucleocapsid protein during human coronavirus 229E replication. *Antiviral Res.* 173, 104620.
- Malbon, A.J., Fonfara, S., Meli, M.L., Hahn, S., Egberink, H., Kipar, A., 2019. Feline infectious peritonitis as a systemic inflammatory disease: contribution of liver and heart to the pathogenesis. *Viruses* 11 (12), 1144.
- Maral, R., Werner, G.H., 1971. Antiviral activity of L-asparaginase. *Nat. New Biol.* 232 (2), 187–188.
- Martinez, M.A., 2020. Compounds with therapeutic potential against novel respiratory 2019 coronavirus. *Antimicrob. Agents Chemother.* 64 (5) (e00399-20).
- Masters, P.S., Perlman, S., 2013. Coronaviridae. In: Knipe, D.M., Howley, P.M. (Eds.), *Fields Virology*, 6th edn. vol 2. Lippincott Williams & Wilkins, Philadelphia, pp. 825–858.
- McAleer, J.P., Kolls, J.K., 2018. Contributions of the intestinal microbiome in lung immunity. *Eur. J. Immunol.* 48, 39–49.
- McCreary, E., Pogue, J., 2020. COVID-19 treatment: a review of early and emerging options. *Open Forum Infectious Diseases*. 7 <https://doi.org/10.1093/ofid/ofaa105>.
- Mehta, P., McAuley, D.F., Brown, M., Sanchez, E., Tattersall, R.S., Manson, J.J., HLH Across Speciality Collaboration, UK, 2020. COVID-19: consider cytokine storm syndromes and immunosuppression. *Lancet* 395 (10229), 1033.
- Miguel, B., Pharr, G.T., Wang, C., 2002. The role of feline aminopeptidase N as a receptor for infectious bronchitis virus. *Brief Review Arch Virol.* 147 (11), 2047–2056.
- Millet, J.K., Whittaker, G.R., 2015. Host cell proteases: critical determinants of coronavirus tropism and pathogenesis. *Virus Res.* 202, 120–134.

- Monti, S., Balduzzi, S., Delvino, P., Bellis, E., Quadrelli, V.S., Montecucco, C., 2020. Clinical course of COVID-19 in a series of patients with chronic arthritis treated with immunosuppressive targeted therapies. *Ann. Rheum. Dis.* 79 (5), 667–668.
- Mosell, E.C., Wang, J., Jeffers, S., Edeen, K.E., Wang, S., Cosgrove, G.P., Funk, C.J., Manzer, R., Miura, T.A., Pearson, L.D., Holmes, K.V., Mason, R.J., 2008. SARS-CoV replicates in primary human alveolar type II cell cultures but not in type I-like cells. *Virology* 372 (1), 127–135.
- Mulangu, S., Dodd, L.E., Davey Jr., R.T., Tshiani Mbaya, O., Proschan, M., Mukadi, D., Lusakibanza Manzo, M., Nzolo, D., Tshomba Oloma, A., Ibanda, A., Ali, R., Coulibaly, S., Levine, A.C., Grais, R., Diaz, J., Lane, H.C., Muyembe-Tamfum, J.J., PALM Writing Group, Sivahera, B., Camara, M., Kojan, R., Walker, R., Dighero-Kemp, B., Cao, H., Mukumbayi, P., Mbala-Kingebeni, P., Ahuka, S., Albert, S., Bonnett, T., Crozier, I., Duvenhage, M., Proffitt, C., Teitelbaum, M., Moench, T., Aboulhab, J., Barrett, K., Cahill, K., Cone, K., Eckes, R., Hensley, L., Herpin, B., Higgs, E., Ledgerwood, J., Pierson, J., Smolskis, M., Sow, Y., Tierney, J., Sivapalasingam, S., Holman, W., Gettinger, N., Vallée, D., Nordwall, J., PALM Consortium Study Team, 2019. A randomized, controlled trial of ebola virus disease therapeutics. *N. Engl. J. Med.* 381 (24), 2293–2303.
- Murphy, B.G., Perron, M., Murakami, E., Bauer, K., Park, Y., Eckstrand, C., Liepnieks, M., Pedersen, N.C., 2018. The nucleoside analog GS-441524 strongly inhibits feline infectious peritonitis (FIP) virus in tissue culture and experimental cat infection studies. *Vet. Microbiol.* 219, 226–233.
- Oostra, M., de Haan, C.A., de Groot, R.J., Rottier, P.J., 2006. Glycosylation of the severe acute respiratory syndrome coronavirus triple-panning membrane proteins 3a and M. *J. Virol.* 80, 2326–2336.
- Ou, X., Liu, Y., Lei, X., Li, P., Mi, D., Ren, L., Guo, L., Guo, R., Chen, T., Hu, J., 2020. Characterization of spike glycoprotein of SARS-CoV-2 on virus entry and its immune cross-reactivity with SARS-CoV. *Nat. Commun.* 11, 1620. <https://doi.org/10.1038/s41467-020-15562-9>.
- Pedersen, N.C., 2009. A review of feline infectious peritonitis virus infection: 1963–2008. *J. Feline Med. Surg.* 11 (4), 225–258.
- Pedersen, N.C., 2014. An update of feline infectious peritonitis: diagnostics and therapeutics. *Vet. J.* 201 (2), 133–141.
- Pedersen, N.C., Black, J.W., Boyle, J.F., Evermann, J.F., McKeirnan, A.J., Ott, R.L., 1984. Pathogenic differences between various feline coronavirus isolates. *Adv. Exp. Med. Biol.* 173, 365–380.
- Pedersen, N.C., Kim, Y., Liu, H., Galasiti Kankanamalage, A.C., Eckstrand, C., Groutas, W. C., Bannasch, M., Meadows, J.M., Chang, K.O., 2018. Efficacy of a 3C-like protease inhibitor in treating various forms of acquired feline infectious peritonitis. *J. Feline Med. Surg.* 20 (4), 378–392.
- Pedersen, N.C., Perron, M., Bannasch, M., Montgomery, E., Murakami, E., Liepnieks, M., Liu, H., 2019. Efficacy and safety of the nucleoside analog GS-441524 for treatment of cats with naturally occurring feline infectious peritonitis. *J. Feline Med. Surg.* 21 (4), 271–281.
- Peters, H.L., Jochmans, D., de Wilde, A.H., Posthuma, C.C., Snijder, E.J., Neyts, J., Seley-Radtke, K.L., 2015. Design, synthesis and evaluation of a series of acyclic fleximer nucleoside analogues with anti-coronavirus activity. *Bioorg. Med. Chem. Lett.* 25, 2923–2926.
- Pfefferle, S., Schöpf, J., Kögl, M., Friedel, C.C., Müller, M.A., Carbajo-Lozoya, J., Stellberger, T., von Dall'Armi, E., Herzog, P., Kallies, S., Niemeyer, D., Ditt, V., Kuri, T., Züst, R., Pumpor, K., Hilgenfeld, R., Schwarz, F., Zimmer, R., Steffen, I., Weber, F., Thiel, V., Herrler, G., Thiel, H.J., Schwegmann-Wessels, C., Pöhlmann, S., Haas, J., Drosten, C., von Brunn, A., 2011. The SARS-coronavirus-host interactome: identification of cyclophilins as target for pan-coronavirus inhibitors. *PLoS Pathog.* 7 (10), e1002331.
- Protzer, U., Seyfried, S., Quasdorff, M., Sassi, G., Svorcova, M., Webb, D., Bohne, F., Hosel, M., Schirmacher, P., Tiegs, G., 2007. Antiviral activity and hepatoprotection by heme oxygenase-1 in hepatitis B virus infection. *Gastroenterology* 133, 1156–1165.
- Pruijssers, A.J., Denison, M.R., 2019. Nucleoside analogues for the treatment of coronavirus infections. *Curr. Opin. Virol.* 35, 57–62.
- Pyrk, K., Bosch, B.J., Berkhout, B., Jebbink, M.F., Dijkman, R., Rottier, P., van der Hoek, L., 2006. Inhibition of human coronavirus NL63 infection at early stages of the replication cycle. *Antimicrob. Agents Chemother.* 50, 2000–2008.
- Rockwell, N.C., Krysan, D.J., Komiyama, T., Fuller, R.S., 2002. Precursor processing by kex2/furin proteases. *Chem. Rev.* 102 (12), 4525–4548.
- Rolain, J.M., Colson, P., Raoult, D., 2007. Recycling of chloroquine and its hydroxyl analogue to face bacterial, fungal and viral infections in the 21st century. *Int. J. Antimicrob. Agents.* 30, 297–308.
- Rose-John, S., 2012. IL-6 trans-signaling via the soluble IL-6 receptor: importance for the pro-inflammatory activities of IL-6. *Int. J. Boil. Sci.* 8, 1237–1247.
- Saijo, M., Morikawa, S., Fukushi, S., Mizutani, T., Hasegawa, H., Nagata, N., Iwata, N., Kurane, I., 2005. Inhibitory effect of mizoribine and ribavirin on the replication of severe acute respiratory syndrome (SARS)-associated coronavirus. *Antiviral Res.* 66, 159–163.
- Sarma, P., Prajapat, M., Avti, P., Kaur, H., Kumar, S., Medhi, B., 2020. Therapeutic options for the treatment of 2019-novel coronavirus: an evidence-based approach. *Indian J. Pharmacol.* 52 (1), 1–5.
- Sarzi-Puttini, P., Giorgi, V., Sirotti, S., Marotto, D., Ardizzone, S., Rizzardini, G., Antinori, S., Galli, M., 2020. COVID-19, cytokines and immunosuppression: what can we learn from severe acute respiratory syndrome? *Clin. Exp. Rheumatol.* 38, 337–342.
- Schmiege, D., Perez Arredondo, A.M., Ntatal, J., Minetto Gellert Paris, J., Savi, M.K., Patel, K., Yasobant, S., Falkenberg, T., 2020. One Health in the context of coronavirus outbreaks: a systematic literature review. *One Health* 100170.
- Schoeman, D., Fielding, B.C., 2019. Coronavirus envelope protein: current knowledge. *Virol. J.* 16, 69.
- Shannon, A., Le, N.T., Selisko, B., Eydoux, C., Alvarez, K., Guillemot, J.C., Decroly, E., Peersen, O., Ferron, F., Canard, B., 2020. Remdesivir and SARS-CoV-2: Structural requirements at both nsp12 RdRp and nsp14 Exonuclease active-sites. *Antivir. Res.* 104793.
- Sheahan, T.P., Sims, A.C., Graham, R.L., Menachery, V.D., Gralinski, L.E., Case, J.B., Leist, S.R., Pyrc, K., Feng, J.Y., Trantcheva, I., Bannister, R., Park, Y., Babusis, D., Clarke, M.O., Mackman, R.L., Spahn, J.E., Palmiotti, C.A., Siegel, D., Ray, A.S., Cihlar, T., Jordan, R., Denison, M.R., Baric, R.S., 2017. Broad-spectrum antiviral GS-5734 inhibits both epidemic and zoonotic coronaviruses. *Sci. Transl. Med.* 9, 396.
- Shiba, N., Maeda, K., Kato, H., Mochizuki, M., Iwata, H., 2007. Differentiation of feline coronavirus type I and II infections by virus neutralization test. *Vet. Microbiol.* 124, 348–352.
- Siddell, S., Ziebuhr, J., Snijder, E.J., 2005. Coronaviruses, toroviruses, and arteriviruses. In: Mahy, B.W.J., Ter Meulen, V. (Eds.), *Topley & Wilson's Microbiology and Microbial Infections, Virology*. Hodder Arnold, London, pp. 823–856.
- Song, H., Seddighzadeh, B., Cooperberg, M.R., Huang, F.W., 2020. Expression of ACE2 and TMPRSS2, the SARS2-CoV-2 receptor and co-receptor, in prostate epithelial cells. *BioRxiv* 2020 (04), 24.056259.
- Stanley, P., Taniguchi, N., Aebi, M., 2017. N-Glycans. In: Varki, A., Cummings, R.D., Esko, J.D., et al. (Eds.), *Essentials of Glycobiology*, 3rd edition. Cold Spring Harbor Laboratory Press, Cold Spring Harbor (NY). Internet. (2015–2017. Chapter 9).
- Sutton, T.C., Subbarao, K., 2015. Development of animal models against emerging coronaviruses: From SARS to MERS coronavirus. *Virology* 479, 247–258.
- Szczepanski, A., Owczarek, K., Milewska, A., Baster, Z., Rajfur, Z., Mitchell, J.A., Pyrc, K., 2018. Canine respiratory coronavirus employs caveolin-1-mediated pathway for internalization to HRT-18G cells. *Vet. Res.* 49, 55.
- The Severe Covid-19 GWAS Group, 2020. Genome-wide association study of severe COVID-19 with respiratory failure. *N. Engl. J. Med.* 383 (16), 1522–1534.
- Triplet, B., Howard, M.W., Jobling, M., Holmes, R.K., Holmes, K.V., Hodges, R.S., 2004. Structural characterization of the SARS-coronavirus spike S fusion protein core. *J. Biol. Chem.* 279, 20836–20849.
- Trompette, A., Gollwitzer, E.S., Yadava, K., Sichelstiel, A.K., Sprenger, N., Ngom-Bru, C., Blanchard, C., Junt, T., Nicod, L.P., Harris, N.L., Marsland, B.J., 2014. Gut microbiota metabolism of dietary fiber influences allergic airway disease and hematopoiesis. *Nat. Med.* 20, 159–166.
- Tseng, C.K., Lin, C.K., Wu, Y.H., Chen, Y.H., Chen, W.C., Young, K.C., Lee, J.C., 2016. Human heme oxygenase 1 is a potential host cell factor against dengue virus replication. *Sci Rep.* 6, 32176.
- van Regenmortel, M.H.V., Fauquet, C.M., Bishop, D.H.L., Carstens, E.B., Estes, M.K., Lemon, S.M., et al., 2000. Coronaviridae. In: Fauquet, C.M., Bishop, D.H.L., Carstens, E.B., Estes, M.K., Lemon, S.M., et al. (Eds.), *Virus taxonomy: Classification and nomenclature of viruses Seventh report of the International Committee on Taxonomy of Viruses*. Academic Press, San Diego, pp. 835–849 (ISBN 0123702003).
- Veronese, N., Demurtas, J., Yang, L., Tonelli, R., Barbagallo, M., Lopalco, P., Lagolio, E., Celotto, S., Pizzol, D., Zou, L., Tully, M.A., Ilie, P.C., Trott, M., López-Sánchez, G.F., Smith, L., 2020. Use of corticosteroids in coronavirus disease 2019 pneumonia: a systematic review of the literature. *Front. Med.* 7, 170.
- Vigerust, D.J., Shepherd, V.L., 2007. Virus glycosylation: role in virulence and immune interactions. *Trends Microbiol.* 15 (5), 211–218.
- Warren, T.K., Wells, J., Panchal, R.G., Stuthman, K.S., Garza, N.L., VanTongeren, S.A., Dong, L., Retterer, C.J., Eaton, B.P., Pegoraro, G., et al., 2014. Protection against filovirus diseases by a novel broad-spectrum nucleoside analogue BCX4430. *Nature* 508, 402–405.
- Warren, T.K., Jordan, R., Lo, M.K., Ray, A.S., Mackman, R.L., Soloveva, V., Siegel, D., Perron, M., Bannister, R., et al., 2016. Therapeutic efficacy of the small molecule GS-5734 against Ebola virus in rhesus monkeys. *Nature* 531 (7594), 381–385.
- Weiss, S.R., Navas-Martin, S., 2005. Coronavirus pathogenesis and the emerging pathogen severe acute respiratory syndrome coronavirus. *Microbiol. Mol. Biol. Rev.* 69, 635–664.
- Wong, C.K., Lam, C.W.K., Wu, A.K.L., 2004. Plasma inflammatory cytokines and chemokines in severe acute respiratory syndrome. *Clin. Exp. Immunol.* 136, 95–103.
- World Health Organization, 2020. SARS (Severe Acute Respiratory Syndrome). World Health Organization (Last accessed on 2020 Feb 13).
- Xiao, X., Chakraborti, S., Dimitrov, A.S., Gramatkov, K., Dimitrov, D.S., 2003. The SARS-CoV S glycoprotein: expression and functional characterization. *Biochem. Biophys. Res. Commun.* 312, 1159–1164.
- Yao, X., Ye, F., Zhang, M., Cui, C., Huang, B., Niu, P., 2020. In vitro antiviral activity and projection of optimized dosing design of hydroxychloroquine for the treatment of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2). *Clin. Infect. Dis.* 71 (15), 732–739.
- Yu, L., Tong, Y., Shen, G., Fu, A., Lai, Y., Zhou, X., Yuan, Y., Wang, Y., Pan, Y., Yu, Z., Li, Y., Liu, T., Jiang, H., 2020. Immunodepletion with hypoxemia: a potential high risk subtype of coronavirus disease 2019. *MedRxiv*. <https://doi.org/10.1101/2020.03.03.20030650>.
- Zeberg, H., Pääbo, S., 2020. The major genetic risk factor for severe COVID-19 is inherited from Neanderthals. *Nature* 587, 610–612.
- Zhou, G., Zhao, Q., 2020. Perspectives on therapeutic neutralizing antibodies against the Novel Coronavirus SARS-CoV-2. *Int. J. Biol. Sci.* 16 (10), 1718–1723.
- Zhu, H., Shi, X., Ju, D., Huang, H., Wei, W., Dong, X., 2014. Anti-inflammatory effect of thalidomide on H1N1 influenza virus-induced pulmonary injury in mice. *Inflammation* 37 (6), 2091–2098.
- Zhu, L., Xu, X., Ma, K., Yang, J., Guan, H., Chen, S., Chen, Z., Chen, G., 2020. Successful recovery of COVID-19 pneumonia in a renal transplant recipient with long-term immunosuppression. *Am. J. Transplant.* 20 (7), 1859–1863.