ORIGINAL PAPER



Green drugs in the fight against *Anisakis simplex*—larvicidal activity and acetylcholinesterase inhibition of *Origanum compactum* essential oil

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Abstract

Anisakiasis is a fish-borne parasitic disease caused by the consumption of raw or undercooked fish, as well as cephalopods, contaminated by third instar larvae (L3) of species belonging to the genus *Anisakis* (Anisakidae). *Origanum compactum* is a small herbaceous aromatic plant endemic to Spain and Morocco. In Morocco, the plant is used under infusion to treat heart diseases and intestinal pains or as preservative for foodstuffs. This is the first time that the *O. compactum* essential oil is tested against the parasitic nematode *Anisakis simplex*. The phytochemical analysis by GC-MS revealed carvacrol (50.3%) and thymol (14.8%) as the major oil constituents. The essential oil and its major constituents carvacrol and thymol were tested against *A. simplex* L3 larvae isolated from blue whiting fish (*Micromesistius poutassou*). *A. simplex* mortality (%) after 24 and 48 h of treatment at 1 µl/ml was 100%, with a low LD₅₀ compared with other essential oils and extracts, and the penetration in the agar assay was also reduced, if compared with control wells. The oil, as well as its major constituents, demonstrated a dose-dependent larvicidal activity. Inhibition of the enzyme acetylcholinesterase through a colorimetric assay in 96-well plates was used to elucidate the pharmacological mechanism as this enzyme plays a key role in nematodes neuromuscular function. Interestingly, *O. compactum* essential oil, carvacrol and thymol inhibited the enzyme, confirming that this could be one of the mechanisms involved in the anthelmintic activity. To the best of our knowledge, this is the first time that *O. compactum* essential oil is reported as a larvicidal agent against *A. simplex* L3 larvae.

Keywords Carvacrol · Thymol · Anisakis · Anthelmintic · Nematode · Waterborne parasitology

Introduction

Anisakiasis is a fish-borne parasitic disease; it is caused by the consumption of raw or undercooked fish, as well as cephalo-

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pods, contaminated by third instar larvae of species belonging to the genus *Anisakis* (Anisakidae), with special reference to *Anisakis simplex*, *Anisakis pegreffii* and *Pseudoterranova decipiens* (Grabda 1976; Mattiucci and Nascetti 2006; Audicana and Kennedy 2008; Pravettoni et al. 2012). After ingestion, the larvae penetrate the gastric and intestinal mucosa, causing the symptoms of anisakiasis (Broglia and Kapel 2011). Recently, it has been estimated that about 20,000 cases of anisakiasis are reported per year worldwide. The large majority of the cases is from Japan (> 90%), due to the traditional use of *sashimi* as food. Other countries characterized by a rather large number of anisakiasis cases are Spain, the Netherlands, Germany and Scandinavia (Bao et al. 2017).

Live larvae of *A. simplex* can cause a parasitic infection of the digestive tract. In addition, they can also affect other organs causing erosive and/or haemorrhagic lesions, ascites and perforations until granulomas and masses, if the larvae are not promptly removed. Furthermore, the presence of larvae of *A. simplex* can lead to allergic reactions, as anaphylaxis, acute/ chronic urticaria and angioedema (Asturias et al. 2000; Nieuwenhuizen et al. 2003; Berger and Marr 2006; Choi et al. 2009; Pravettoni et al. 2012). Notably, the larvae of *A. simplex* can induce an immune adaptive response characterized by T-lymphocyte proliferation with polyclonal and monoclonal (which are responsible for allergic symptoms), IgE production, eosinophilia and mastocytosis (Park et al. 2009; Pravettoni et al. 2012).

Nowadays, the endoscopic removal of live larvae still represents the main effective treatment against anisakiasis (Sugita et al. 2008; Pravettoni et al. 2012). Furthermore, protection against anisakiasis can be enhanced educating consumers about the risks linked with eating raw or undercooked seafood. From a pharmacological point of view, antibiotics, anticholinergics and/or corticosteroids have been employed for the treatment of anisakiasis, showing patchy and limited efficacy (Matsui et al. 1985). However, a relevant exception is represented by albendazole (Dziekońska-Rynko et al. 2002; Romero et al. 2014), since it has been recently showed that this compound dose-dependently reduced (500 µg/ml led to 100% 48 h post-treatment) the survival of A. simplex third instar larvae (Arias-Diaz et al. 2006). However, it has been noted that acidic medium pH reduced its efficacy (Arias-Diaz et al. 2006). In this scenario, there is an urgent need to develop novel and effective tools and drugs for the treatment of A. simplex parasitic infections (Molina-Garcia and Sanz 2002; Brutti et al. 2010), including the use of natural products (Hierro et al. 2004, 2006; Lin et al. 2010; Gómez-Rincón et al. 2014; Valero et al. 2015).

Essential oils, which are complex mixtures of volatile components such as monoterpenoids and sesquiterpenoids (Pavela and Benelli 2016), can represent a new strategy to combat several parasites including L3 larvae of *A. simplex* (Giarratana et al. 2014; Hierro et al. 2004; Romero et al. 2012). In this regard, *Origanum* essential oils have been shown to possess in vivo efficacy against larvae of *Anisakis* (Pérez et al. 2016; Abattouy et al. 2010). On this basis, we focused our attention on the essential oil obtained from *Origanum compactum* Benth., belonging to the Lamiaceae family.

O. compactum is a small herbaceous aromatic plant endemic to Spain and Morocco (Emberger and Maire 1941) where it is particularly appreciated in cuisine to enhance the flavour of foods and as natural food preservative (Ghanmi et al. 2015; Sbayou et al. 2014). *O. compactum* enjoys a long-standing use in the traditional medicine as well as a good reputation and economic importance in the trade of medicinal and aromatic plants (Briguiche and Zidane 2016; Bouiamrine et al. 2017). In Morocco, the plant, locally known as *Zaetar*, is used under infusion to treat heart and intestinal pains or as

preservative for foodstuffs (El-Hilaly et al. 2003). It is also a famous remedy for the treatment of wounds, diabetes, hypertension and cardiac diseases, digestive and respiratory problems and gastrointestinal and gingival cancers (Ziyyat et al. 1997; Jouad et al. 2001; Kabbaj et al. 2012; Eddouks et al. 2002; Bouhdid et al. 2009; Jamila and Mostafa 2014). *O. compactum* has also been used as vermifuge, aphrodisiac, antispasmodic, antiulcer, laxative and antidiarrheal agent (Hmamouchi et al. 2000; Jamila and Mostafa 2014).

In the search of scalable products to use as treatments of anisakiasis, we here evaluated the activity of *O. compactum* essential oil as well as its major constituents carvacrol and thymol on *A. simplex* L3 larvae. The pharmacological mechanism was studied through the inhibitory activity of the acetylcholinesterase enzyme, which plays a crucial role in the nematode neuromuscular function.

Materials and methods

Origanum compactum essential oil

The essential oil of O. compactum (obtained from flowering aerial parts) was kindly supplied by Pranarôm International (http://www.pranarom.com). Its analysis was achieved on an Agilent 6890 N gas chromatograph equipped with a 5973-N mass spectrometer (MS). The settings for the MS were as follows: EI mode, 70 eV, and mass to charge ratio (m/z) scan between 35 and 400. A HP-5 MS capillary column (30 m × ID 0.25 mm × 0.25 µm film thickness, J & W Scientific, Folsom, CA, USA) using helium gas flow (1.0 ml/min) was used for separation. The GC temperature program was as follows: initial 50 °C for 5 min, then increasing with 20 °C/min to 300 °C. The injector temperature was 150 °C. Qualiquantitative analysis of the essential oil was performed according to the work of Benelli et al. (2017). Thymol and carvacrol were purchased from Sigma-Aldrich (Madrid, Spain).

Anisakis simplex from blue whiting

Anisakis simplex L3 larvae were isolated from the intermediary host *Micromesistius poutassou* (Risso) (blue whiting) purchased from the fishmonger in Villanueva de Gállego (Zaragoza, Spain). The worms were washed several times on sterile solution of 0.9% NaCl and identified under light microscope according to morphological features. Only larvae with length > 2.0 cm were used for the larvicidal assays.

Anthelmintic activity against Anisakis simplex L3 larvae

This assay was performed in 6 well plates. Ten larvae were introduced in each well of polystyrene plates with a final volume of 2 ml sterile saline solution containing different concentrations of the test solution (Gómez-Rincón et al. 2014). *O. compactum* essential oil was tested against *Anisakis* in the range of $0-1 \mu$ l/ml. The major constituents carvacrol and thymol were tested in the same range of concentrations. Appropriate control wells without treatments were also carried out in each experiment. The parasites were incubated at 37 °C for 24 and 48 h. Levamisole was used as the reference antiparasitic drug. Larvae were examined at 24 and 48 h under microscope, and immobile L3 was considered dead.

Inhibition of acetylcholinesterase

The inhibition of acetylcholinesterase (AChE) was determined in 96 microplates using the method by Ellman et al. (1961) with some modifications. Each well contained 25 μ l of 15 mM ATCI in Millipore water, 125 μ l of 3 mM DTNB in buffer C (50 mM Tris–HCl, pH 8, 0.1 M NaCl, 0.02 M MgCl2 6 H2O), 50 μ l buffer B (50 mM Tris–HCl, pH 8, 0.1% bovine serum), and 25 μ l of test compound or extract. Every concentration was tested in triplicates. Then, 25 μ l and 0.22 U/ml AChE were added and the absorbance was measured eight times every 13 s at 405 nm. Galantamine was used as reference.

Penetration assays

Agar block plates were prepared in 12-well plates to study the capacity of the living larvae to penetrate. The agar solution was made with the following reagents: 0.75% agar in RPMI 1640 medium solution (pH 4, Sigma, USA) with 20% FBS (Lonza, USA). One millilitre of the solution was poured into each well. Then, 100 μ l of supernatant, RPMI-1640 (RPMI-1640, 20% FBS, 1% commercial pepsin, pH 4.0), was placed into each well. *A. simplex* L3 larvae were incubated with a sub-lethal concentration (0.125 mg/ml) of *O. compactum* essential oil for 24 h. Five larvae were placed on each control or simple well. The plates were incubated for 24 h, and the number of L3 larvae that penetrated the solid agar block was counted after that period.

Statistical analysis

All experiments were performed in triplicates in different weeks using new *A. simplex* larvae. LD_{50} (median lethal dose) and IC_{50} (half maximal inhibitory concentration) values for

larvicidal and mechanistic assays were calculated using nonlinear regression (GraphPad Prism 5).

Results

Essential oil composition

O. compactum essential oil was characterized by eight chemical compounds accounting for 93.6% of the total composition. The major component was the monoterpene phenol carvacrol accounting for half essential oil composition (50.32%), followed by its isomer thymol (14.8%) and by γ -terpinene (13.6%) and *p*-cymene (8.40%). Minor components were (*E*)-caryophyllene (2.1%), α -terpinene (1.6%), myrcene (1.5%) and linalool (1.3%).

Anthelmintic activity

O. compactum essential oil showed a dose dependent larvicidal activity at 24 and 48 h of treatments. All larvae were killed at doses of 1 μ l/ml after 24 h showing a paralysis that indicates mortality. Although the efficacy of the treatment can be considered similar at 24 and 48 h because the mortality was 100% (Fig. 1), the LD₅₀ of essential oil was lower after 48 h.

The effects of carvacrol and thymol on the larvae were similar to those of the oil (Fig. 2); however, carvacrol exhibited a stronger activity than *O. compactum* and thymol, indicating that this compound might be responsible for the larvicidal effects. Levamisole induced 100% mortality at 0.1 mg/ml.

Inhibition of acetylcholinesterase

O. compactum essential oil, carvacrol and thymol acted as AChE inhibitors, indicating that this could be one of the mechanisms involved in the larvicidal activity (Fig. 3). The activity profile of carvacrol on the enzyme was very similar to that of the essential oil. However, higher doses of thymol were needed to inhibit the AChE enzyme at the same level. The AChE inhibitory potential decreased in the following order: carvacrol > *O. compactum* > thymol. Galantamine at 0.1 mg/ml induced 100% of AChE inhibition.

Penetration assays

The penetration ability of *A. simplex* L3 larvae was evaluated after exposure to *O. compactum* essential oil and compared with untreated larvae. The penetration rate of control wells was constant at 100% while it decreased 25% for treated larvae with sub-lethal doses of the oil (the agar was penetrated by 75% of the larvae).

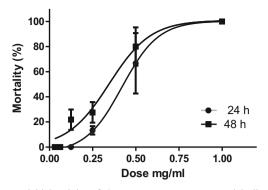


Fig. 1 Larvicidal activity of *Origanum compactum* essential oil against *Anisakis simplex* L3 larvae after 24 and 48 h (LD_{50} 0.429 mg/ml at 24 h and 0.344 mg/ml at 48 h)

Discussion

In the present work, the essential oil from *O. compactum* was assayed for the first time against L3 larvae of *A. simplex*. For the purpose, a commercial oil was used and analysed showing a chemical composition fully consistent with those reported in literature concerning Moroccan oregano accessions, with phenolic compounds such as carvacrol and thymol as the most abundant components, followed by their biogenetic precursors γ -terpinene and *p*-cymene (Bakhy et al. 2014)

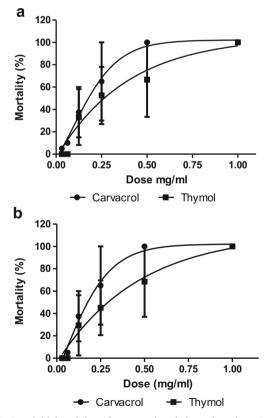


Fig. 2 Larvicidal activity of carvacrol and thymol against *Anisakis simplex* L3 larvae at 24 (**a**) and 48 h (**b**). LD_{50} values for carvacrol were 0.176 mg/ml at 24 h and 0.178 mg/ml at 48 h. LD_{50} values for thymol were 0.291 mg/ml at 24 h and 0.214 mg/ml at 48 h

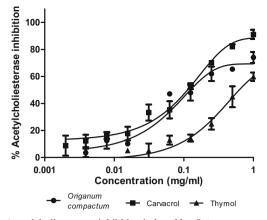


Fig. 3 Acetylcholinesterase inhibition induced by *Origanum compactum* essential oil (IC_{50} 0.124 mg/ml), carvacrol (IC_{50} 0.113 mg/ml) and thymol (IC_{50} 0.625 mg/ml)

(Table 1). Only quantitative differences have hitherto noticed among the main essential oil constituents. Notably, the monoterpene phenols thymol and carvacrol may be detected in comparable amounts (Ghanmi et al. 2015; Kloucek et al. 2012; Sbayou et al. 2014; Bouhdid et al. 2008; Mezzoug et al. 2007) or with the latter more abundant than the former (Bouchra et al. 2003; Bakhy et al. 2014; Pavela 2008; Lamiri et al. 2001) as in our case. Thus, our sample of *O. compactum* can be defined as a carvacrol-rich chemotype (Bakhy et al. 2014).

In our experiments, *O. compactum* essential oil showed anthelmintic effects against *A. simplex* L3 larvae as well as inhibitory activity of the acetylcholinesterase enzyme. Levamisole is used as an antiparasitic agent in veterinary, so we tried the concentration of 0.1 mg/ml as a control substance. One microlitre per millilitre of *O. compactum* essential oil is equivalent to 1 mg/ml approximately, so levamisole was more potent. However, this treatment is not available for humans and is used only in veterinary.

To the best of our knowledge, these results are new and here published for the first time. On the other hand, the antimicrobial activities of this oil were already known and previously reported. O. compactum essential oil is considered as a strong bactericidal and fungicidal, being capable to inhibit the growth of several pathogenic strains such as Staphylococcus aureus, Pseudomonas aeruginosa, Listeria spp., Alternaria alternata and Aspergillus niger (Bouhdid et al. 2009; Kloucek et al. 2012). The antimicrobial effects are related to the presence of monoterpene phenols thymol and carvacrol which can alter permeability of cell membrane and destabilize respiratory and enzymatic activities (Bouyahya et al. 2017; Bakkali et al. 2008). O. compactum essential oil has also shown toxic effects on Botrytis cinerea (Bouchra et al. 2003) as well as on wood decay fungi (Ghanmi et al. 2015), nematicidal activity against Ditylenchus dipsaci (Zouhar et al. 2009) and insecticidal effects against larvae of Spodoptera littoralis (Pavela 2005) and adults of Musca domestica and

Table 1	Chemical composition of the	Origanum compactum	essential oil tested against A	nisakis simplex L3 larvae
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Number	Component ^a	RI Exp. ^b	RI ADAMS ^c	Percent ^d	ID ^e
1	Myrcene	983	988	$1.5 \pm 0.2\%$	RI, MS
2	α-Terpinene	1010	1014	$1.6\pm0.2\%$	RI, MS
3	<i>p</i> -Cymene	1017	1020	$8.4\pm1.1\%$	RI, MS
4	γ-Terpinene	1051	1054	$13.6\pm2.1\%$	RI, MS
5	Linalool	1099	1095	$1.3\pm0.2\%$	RI, MS
6	Thymol	1294	1289	$14.8\pm2.8\%$	Std, RI, MS
7	Carvacrol	1299	1298	$50.3\pm3.9\%$	Std, RI, MS
8	(E)-caryophyllene	1410	1417	$2.1\pm0.4\%$	
	Total identified (%)			93.6%	
	Grouped components (%)				
	Monoterpene hydrocarbons			25.1	
	Oxygenated monoterpene	s		66.4	
	Sesquiterpene hydrocarbo	ons		2.1	

^a Compounds are listed in order of their elution from a HP-5MS column

^b Linear retention index on HP-5MS column, experimentally determined using homologous series of C₈-C₃₀ alkanes

^c Linear retention index taken from Adams (2007)

^d Percentage values are means of three independent analyses \pm SD

^e Identification methods: Std., based on comparison with authentic compounds; MS, based on comparison with ADAMS, FFNSC 2 (2012) and NIST 08 (2008) MS databases; RI, based on comparison of RI with those reported in ADAMS, FFNSC 2 (2012) and NIST 08 (2008)

Mayetiola destructor (Pavela 2008; Lamiri et al. 2001). The ethyl acetate extract was highly active against the schistosomiasis-transmitting snail *Bulinus truncatus* (Hmamouchi et al. 2000).

With the aim of elucidating which compounds of O. compactum essential oil were responsible for the anti-Anisakis activities, the two main monoterpenes, carvacrol and thymol, were also assayed. According to our data, carvacrol might be one of the active principles as it exhibited a higher larvicidal activity than thymol and the oil, revealing its potential as an anthelmintic drug. This is not the first time that carvacrol has been evaluated as a nematicidal compound (Trailović et al. 2015; Andre et al. 2016); in fact, the larvicidal activity of this monoterpene has also been evaluated on A. simplex (Hierro et al. 2004). The inhibitory activity of carvacrol and thymol on the acetylcholinesterase enzyme has also been tested (Jukic et al. 2007). Carvacrol and thymol are isomeric phenolic monoterpenes present in several aromatic medicinal and culinary plants. The different position of the hydroxyl group in the phenyl ring provides better anthelmintic and anti-cholinesterase activities for carvacrol in our case, which agrees with results of other authors (Jukic et al. 2007; Aazza et al. 2011; Seo et al. 2015).

Overall, the results of the present study pointed out the importance of the traditional use of *O. compactum* as a food preservative in Morocco and Spain. Its demonstrated anthelmintic properties, together with its well-known antibacterial and fungicidal effects, make it an ideal candidate as

preservative agent for the prevention and treatment of foodborne diseases.

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Compliance with ethical standards

Conflict of interest The authors declare that they have no competing interests.

References

- Aazza S, Lyoussi B, Miguel MG (2011) Antioxidant and antiacetylcholinesterase activities of some commercial essential oils and their major compounds. Molecules 16:7672–7690
- Abattouy N, Valero A, Romero MC, Martin-Sanchez J, Gonzalez-Tejero MR, Lozano J, Navarro MC (2010) *In vivo* activity of essential oil *Origanum elongatum* against larva L3 of *Anisakis pegreffii*. Ars Pharma 51:107–111
- Adams RP (2007) Identification of essential oil components by gas chromatography/mass spectrometry. Allured Publishing Corporation, Carol Stream
- Andre WP, Ribeiro WL, Cavalcante GS, dos Santos JM, Macedo IT, de Paula HC, de Freitas RM, de Morais SM, de Melo JV, Bevilaqua CM (2016) Comparative efficacy and toxic effects of carvacryl acetate and carvacrol on sheep gastrointestinal nematodes and mice. Vet Parasitol 218:52–58

- Arias-Diaz J, Zuloaga J, Vara E, Balibrea J, Balibrea JL (2006) Efficacy of albendazole against *Anisakis simplex* larvae in vitro. Dig Liver Dis 38:24–26
- Asturias JA, Eraso E, Moneo I, Martinez A (2000) Is tropomyosin an allergen in *Anisakis*? Allergy 55:898–898
- Audicana MT, Kennedy MW (2008) Anisakis simplex: from obscure infectious worm to inducer of immune hypersensitivity. Clin Microbiol Rev 21:360–379
- Bakhy K, Benlhabib O, Bighelli A, Casanova J, Tomi F, Al Faiz C (2014) Yield and chemical variability of the essential oil isolated from aerial parts of wild *Origanum compactum* Benth. from Moroccan Western Rif. Am J Essent Oil Nat Prod 1:9–17
- Bakkali F, Averbeck S, Averbeck D, Idaomar M (2008) Biological effects of essential oils—a review. Food Chem Toxicol 46:446–475
- Bao M, Pierce GJ, Pascual S, González-Muñoz M, Mattiucci S, Mladineo I, Cipriani P, Buselic I, Strachan NJC (2017) Assessing the risk of an emerging zoonosis of worldwide concern: anisakiasis. Sci Rep 7: 43699. https://doi.org/10.1038/srep43699
- Benelli G, Pavela R, Iannarelli R, Petrelli R, Cappellacci L, Cianfaglione K, Afshar FH, Nicoletti M, Canale A, Maggi F (2017) Synergized mixtures of Apiaceae essential oils and related plant-borne compounds: larvicidal effectiveness on the filariasis vector *Culex quinquefasciatus* Say. Ind Crops Prod 96:186–195
- Berger SA, Marr JS (2006) Human parasitic diseases sourcebook. Jones and Bartlett Publishers, Sudbury
- Bouchra C, Achouri M, Hassani LMI, Hmamouchi M (2003) Chemical composition and antifungal activity of essential oils of seven Moroccan Labiatae against *Botrytis cinerea* Pers. Fr J Ethnopharmacol 89:165–169
- Bouhdid S, Abrini J, Zhiri A, Espuny MJ, Manresa A (2009) Investigation of functional and morphological changes in *Pseudomonas* aeruginosa and Staphylococcus aureus cells induced by Origanum compactum essential oil. J Appl Microbiol 106:1558–1568
- Bouhdid S, Skali SN, Idaomar M, Zhiri A, Baudoux D, Amensour M, Abrini J (2008) Antibacterial and antioxidant activities of *Origanum compactum* essential oil. Afr J Biotech 7:1563–1570
- Bouiamrine EH, Bachiri L, Ibijbijen J, Nassiri L (2017) Fresh medicinal plants in middle atlas of Morocco: trade and threats to the sustainable harvesting. J Med Plants Studies 5:123–128
- Bouyahya A, Bakri Y, El Ouardy K, Edaoudi F, Talbaoui A, Et-Touys A, Abrini J, Dakka N (2017) Antibacterial, antioxidant and antitumor properties of Moroccan medicinal plants: a review. Asian Pac J Trop Dis 7:57–64
- Briguiche H, Zidane L (2016) Ethnobotanical study of medicinal plants from El-Jadida City (Morocco). Lazaroa 37:145–151
- Broglia A, Kapel C (2011) Changing dietary habits in a changing world: emerging drivers for the transmission of foodborne parasitic zoonoses. Vet Parasitol 182:2–13
- Brutti A, Rovere P, Cavallero S, D'Amelio S, Danesi P, Arcangeli G (2010) Inactivation of *Anisakis simplex* larvae in raw fish using high hydrostatic pressure treatments. Food Control 21:331–333
- Choi SJ, Lee JC, Kim MJ, Hur GY, Shin SY, Park HS (2009) The clinical characteristics of *Anisakis* allergy in Korea. Korean J Intern Med 24: 160–163
- Dziekońska-Rynko J, Rokicki J, Jabłonowski Z (2002) Effects of ivermectin and albendazole against *Anisakis simplex* in vitro and in guinea pig. J Parasitol 88:395–398
- Eddouks M, Maghrani M, Lemhadri A, Ouahidi M-L, Jouad H (2002) Ethnopharmacological survey of medicinal plants used for the treatment of diabetes mellitus, hypertension and cardiac diseases in the south-east region of Morocco (Tafilalet). J Ethnopharmacol 82:97–103
- El-Hilaly J, Hmammouchi M, Lyoussi B (2003) Ethnobotanical studies and economic evaluation of medicinal plants in Taounate province (Northern Morocco). J Ethnopharmacol 86:149–158
- Ellman GL, Courtney KD, Andres V, Featherstone RM (1961) A new and rapidcolorimetric determination of acetylcholinesterase activity. Biochem Pharmacol:88–95

- Emberger L, Maire R (1941) Catalogue of plants of Morocco (Spermatophytes and Pteridophytes). Tome IV, Supplement to volumes I, II, and III. Pp. 915-1181. Minerva, Algiers
- FFNSC 2 (2012) Flavors and fragrances of natural and synthetic compounds. Mass Spectral Database Japan: Shimadzu Corps
- Ghanmi M, Satrani Thevenon MF, Elyounssi K, Ajjourri M (2015) Essential oils from *Origanum compactum* as an alternative active ingredient against wood decay fungi. Nat Prod 11:135–142
- Giarratana F, Muscolino D, Beninati C, Giuffrida A, Panebianco A (2014) Activity of *Thymus vulgaris* essential oil against *Anisakis larvae*. Exp Parasitol 142:7–10
- Gómez-Rincón C, Langa E, Murillo P, Valero MS, Berzosa C, López V (2014) Activity of tea tree (Melaleuca alternifolia) essential oil against L3 larvae of *Anisakis simplex*. Biomed Res Int 2014: 549510. https://doi.org/10.1155/2014/549510
- Grabda J (1976) Studies on the life cycle and morphogenesis of *Anisakis simplex* (Rudolphi, 1809) (Nematoda: Anisakidae) cultured *in vitro*. Acta Ichthyol Piscat 6:119–131
- Hierro I, Valero A, Navarro MC (2006) *In vivo* larvicidal activity of monoterpenic derivatives from aromatic plants against L 3 larvae of *Anisakis simplex* sl. Phytomedicine 13:527–531
- Hierro I, Valero A, Perez P, Gonzalez P, Cabo MM, Montilla MP, Navarro MC (2004) Action of different monoterpenic compounds against *Anisakis simplex* sl L3 larvae. Phytomedicine 11:77–82
- Hmamouchi M, Lahlou M, Agoumi A (2000) Molluscicidal activity of some Moroccan medicinal plants. Fitoterapia 71:308–314
- Jamila F, Mostafa E (2014) Ethnobotanical survey of medicinal plants used by people in oriental Morocco to manage various ailments. J Ethnopharmacol 154:76–87
- Jouad H, Haloui M, Rhiouani H, El Hilaly J, Eddouks M (2001) Ethnobotanical survey of medicinal plants used for the treatment of diabetes, cardiac and renal diseases in the North centre region of Morocco (Fez–Boulemane). J Ethnopharmacol 77:175–182
- Jukic M, Politeo O, Maksimovic M, Milos M, Milos M (2007) In vitro acetylcholinesterase inhibitory properties of thymol, carvacrol and their derivatives thymoquinone and thymohydroquinone. Phytother Res 21:259–261
- Kabbaj FZ, Meddah B, Cherrah Y, Faouzi MEA (2012)
 Ethnopharmacological profile of traditional plants used in Morocco by cancer patients as herbal therapeutics.
 Phytopharmacology 2:243–256
- Kloucek P, Smid J, Frankova A, Kokoska L, Valterova I, Pavela R (2012) Fast screening method for assessment of antimicrobial activity of essential oils in vapor phase. Food Res Int 47:161–165
- Lamiri A, Lhaloui S, Benjilali B, Berrada M (2001) Insecticidal effects of essential oils against Hessian fly, *Mayetiola destructor* (Say). Field Crops Res 71:9–15
- Lin RJ, Chen CY, Lee JD, Lu CM, Chung LY, Yen CM (2010) Larvicidal constituents of *Zingiber officinale* (ginger) against *Anisakis simplex*. Planta Med 76:1852–1858
- Matsui T, Lida M, Murakami M, Kimura Y, Fujishima M, Yao Y, Tsuji M (1985) Intestinal anisakiasis: clinical and radiologic features. Radiology 157:299–302
- Mattiucci S, Nascetti G (2006) Molecular systematics, phylogeny and ecology of anisakid nematodes of the genus *Anisakis* Dujardin, 1845: an update. Parasite 13:99–113
- Mezzoug N, Elhadri A, Dallouh A, Amkiss S, Skali NS, Abrini J, Zhiri A, Baudoux D, Diallo B, El Jaziri M, Idaomar M (2007) Investigation of the mutagenic and antimutagenic effects of *Origanum compactum* essential oil and some of its constituents. Mutat Res 629:100–110
- Molina-Garcia AD, Sanz PD (2002) *Anisakis simplex* larva killed by high-hydrostatic-pressure processing. J Food Prot 65:383–388
- Nieuwenhuizen N, Lopata AL, Jeebhay MF, Herbert DR, Robins TG, Brombacher F (2003) Exposure to the fish parasite *Anisakis* causes

allergic airway hyperreactivity and dermatitis. J Allergy Clin Immunol 117:1098–1105

- NIST 08 (2008) Mass spectral library (NIST/EPA/NIH). National Institute of Standards and Technology, Gaithersburg
- Park SK, Cho MK, Park HK, Lee KH, Lee SJ, Choi SH, Ock MS, Jeong HJ, Lee MH, Yu HS (2009) Macrophage migration inhibitory factor homologs of *Anisakis simplex* suppress Th2 response in allergic airway inflammation model via CD4+ CD25+ Foxp3+ T cell recruitment. J Immunol 182:6907–6914
- Pavela R (2005) Insecticidal activity of some essential oils against larvae of Spodoptera littoralis. Fitoterapia 76:691–696
- Pavela R (2008) Insecticidal properties of several essential oils on the house fly (*Musca domestica* L.) Phytother Res 22:274–278
- Pavela R, Benelli G (2016) Essential oils as eco-friendly biopesticides? Challenges and constraints. Trends Plant Sci 21:1000–1007
- Pérez MGM, Moll CN, Espinosa GM, López AV (2016) Evaluation of different Mediterranean essential oils as prophylactic agents in anisakidosis. Pharm Biol 55:456–461
- Pravettoni V, Primavesi L, Piantanida M (2012) *Anisakis simplex*: current knowledge. Eur Ann Allergy Clin Immunol 44:150–156
- Romero MC, Navarro MC, Martín-Sánchez J, Valero A (2014) Peppermint (*Mentha piperita*) and albendazole against anisakiasis in an animal model. Tropical Med Int Health 19:1430–1436
- Romero MC, Valero A, Martín-Sánchez J, Navarro-Moll MC (2012) Activity of *Matricaria chamomilla* essential oil against anisakiasis. Phytomedicine 19:520–523

- Seo SM, Jung CS, Kang J, Lee HR, Kim SW, Hyun J, Park IK (2015) Larvicidal and acetylcholinesterase inhibitory activities of apiaceae plant essential oils and their constituents against *Aedes albopictus* and formulation development. J Agric Food Chem 63:9977–9986
- Sbayou H, Oubrim N, Bouchrif B, Ababou B, Boukachabine K, Amghar S (2014) Chemical composition and antibacterial activity of essential oil of *Origanum compactum* against foodborne bacteria. Int J Eng Res Technol 3:3562–3567
- Sugita S, Sasaki A, Shiraishi N, Kitano S (2008) Laparoscopic treatment for a case of ileal anisakiasis. Surg Laparosc Endosc Percutan Tech 18:216–218
- Trailović SM, Marjanović DS, Nedeljković Trailović J, Robertson AP, Martin RJ (2015) Interaction of carvacrol with the Ascaris suum nicotinic acetylcholine receptors and gamma-aminobutyric acid receptors, potential mechanism of antinematodal action. Parasitol Res 114:3059–3068
- Valero A, Romero MC, Gómez-Mateos M, Hierro I, Navarro MC (2015) Natural products: perspectives in the pharmacological treatment of gastrointestinal anisakiasis. Asian Pac J Trop Med 8:612–617
- Ziyyat A, Legssyer A, Mekhfi H, Dassouli A, Serhrouchni M, Benjelloum W (1997) Phytotherapy of hypertension and diabetes in oriental Morocco. J Ethnopharmacol 58:45–54
- Zouhar M, Douda O, Lhotsky D, Pavela R (2009) Effect of plant essential oils on mortality of the stem nematode (*Ditylenchus dipsaci*). Plant Prot Sci 45:66–73